



UNIVERSIDADE ESTADUAL DE SANTA CRUZ
PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA E
CONSERVAÇÃO DA BIODIVERSIDADE

ABANDONED EUCALYPTUS PLANTATIONS CAN BE USED AS
CATALYSTS FOR NATURAL REGENERATION

MATHEUS CARVALHO DOS SANTOS

ILHÉUS - BAHIA

2023

MATHEUS CARVALHO DOS SANTOS

**ABANDONED EUCALYPTUS PLANTATIONS CAN BE USED AS
CATALYSTS FOR NATURAL REGENERATION**

Dissertação apresentada à Universidade Estadual de Santa Cruz, como requisito para obtenção do título de Mestre em Ecologia e Conservação da Biodiversidade.

Orientador: Prof. Dr. Luiz Fernando Silva Magnago

ILHÉUS - BAHIA

2023

S237 Santos, Matheus Carvalho dos.
Abandoned eucalyptus plantations can be used as catalysts for natural regeneration / Matheus Carvalho dos Santos. - Ilhéus : UESC, 2023.

[52]f. : il.

Orientador : Luiz Fernando Silva Magnago.

Dissertação (Mestrado) – Universidade Estadual de Santa Cruz. Programa de Pós-graduação em Ecologia e Conservação da Biodiversidade.

Inclui referências.

1. Biodiversidade – Conservação. 2. Florestas – Reprodução – Brasil. 3. Mata atlântica – Conservação. 4. Eucalipto. 5. Ecologia das florestas tropicais. 6. Ecossistema – Recuperação. I. Magnago, Luiz Fernando Silva. II. Título.

CDD – 363.7

Agradeço aos meus pais e a minha família, pelo amor, incentivo e apoio durante a minha vida.

Agradeço ao meu orientador, Luiz Magnago, por todo apoio, pela disponibilidade e orientação, e pelo incentivo durante este período.

Agradeço à professora Ana Paula L. do Couto Santos, por ter participado da minha formação inicial, por ter me acompanhado durante o estágio de docência e pelo constante apoio e incentivo.

Agradeço à Nathalia Safar, pela disponibilidade do banco de dados utilizado neste trabalho.

Agradeço aos amigos e aos colegas de turma, pela convivência e companheirismo em todos os momentos.

Agradeço a UESC, ao PPGECB e ao seu corpo docente, por fazerem parte da minha formação.

O presente trabalho foi realizado com apoio da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Código de Financiamento 001.

SUMÁRIO

RESUMO	6
ABSTRACT	7
INTRODUÇÃO GERAL	8
REFERÊNCIAS	9
Abandoned Eucalyptus plantations can be used as catalysts for natural regeneration	14
Abstract	15
Introduction	16
Materials and methods	18
<i>Study area</i>	18
<i>Data collection</i>	19
<i>Measures of phylogenetic diversity and structure</i>	20
<i>Functional trait matrix</i>	21
<i>Phylogenetic signal of the functional traits</i>	22
<i>Data analysis</i>	22
Results	23
<i>Does Eucalyptus act as an environmental filter?</i>	23
<i>Does phylogenetic diversity increase over succession?</i>	26
<i>Are functional traits conserved across lineages?</i>	28
Discussion	28
<i>Does Eucalyptus act as an environmental filter?</i>	29
<i>Does phylogenetic diversity increase with succession?</i>	30
<i>Are functional traits conserved across lineages?</i>	32
References	33
Data sources	42
SUPPLEMENTARY MATERIAL	47

RESUMO

A Mata Atlântica tem um elevado potencial de regeneração natural em áreas abandonadas sob plantios de eucaliptos. Hipotetizamos que a regeneração natural nas florestas tropicais começa com comunidades compostas por algumas espécies pioneiras, que exibem menor riqueza, diversidade e complexidade do que as comunidades de clímax encontradas no final da sucessão. Aqui, abordamos a diversidade e a estrutura filogenética nas florestas sob plantios abandonados de eucalipto e investigamos se o eucalipto atua como um filtro ambiental que seleciona as espécies mais aparentadas. Comparamos a diversidade (PD, MPD, MNTD) e a estrutura filogenética (sesPD, NRI, NTI) em áreas de regeneração natural da Mata Atlântica em dois tratamentos: i) tratamento misto, espécies nativas sob plantações abandonadas de eucalipto; ii) tratamento nativo, com a presença de espécies nativas. Estabelecemos 17 transectos no tratamento misto e 35 no tratamento nativo. Em cada transecto, amostramos 5-15 parcelas contíguas de 10 m x 10 m. Dentro de cada parcela, amostramos todos os indivíduos vivos com um diâmetro à altura do peito ≥ 10 cm. Não encontramos diferenças significativas entre tratamentos para nenhum dos índices de diversidade filogenética testados. As médias de NRI e NTI foram positivas, indicando agrupamento filogenética em ambos os tratamentos. Nenhum dos índices de diversidade filogenética e de estrutura filogenética foi significativamente afetado pelo aumento da área basal de acordo com os nossos modelos. As plantações de restauração têm objetivos que vão para além da restauração da cobertura florestal a uma área, assegurando a manutenção de espécies nativas evolutivamente únicas, e mantendo serviços e funções essenciais do ecossistema. Ao longo da sucessão, a diversidade filogenética aumentou em ambos os tratamentos sem diferenças significativas. Concluimos que os planos de restauração podem utilizar o eucalipto como ponto de partida para os planos de restauração, especialmente quando o sequestro de carbono juntamente com a biodiversidade é o principal objectivo do projecto de restauração.

Palavras-chave: Conservação da Biodiversidade; Diversidade e Estrutura Filogenética; Mata Atlântica; Plantações Florestais; Restauração Ecológica; Sinal filogenético de traços funcionais; Sub-bosque de Eucalipto; Sucessão secundária.

ABSTRACT

After abandonment, the Atlantic Forest has a high potential for natural regeneration in areas under eucalyptus plantations. We hypothesize that natural regeneration in tropical forests begins with communities composed of a few pioneer species, which exhibit lower richness, diversity, and complexity than climax communities found at the end of succession. Here, we address diversity and phylogenetic structure in eucalyptus understory forests and investigate whether eucalyptus acts as an environmental filter that selects for more phylogenetically related species. We compared diversity (PD, MPD, MNTD) and phylogenetic structure (sesPD, NRI, NTI) in natural regeneration areas of the Atlantic Forest in two treatments: i) mixed treatment, native species under abandoned eucalyptus plantations; ii) native treatment, with the presence of native species. We established 17 transects in the mixed treatment and 35 in the native treatment. On each transect, we sampled 5-15 contiguous 10 m x 10 m plots. Within each plot, we sampled all live individuals with a diameter at breast height ≥ 10 cm. We found no significant differences between treatments for any of the phylogenetic diversity indices tested. The means of NRI and NTI were positive, indicating phylogenetic clustering in both treatments. None of the phylogenetic diversity and phylogenetic structure metrics were significantly affected by increasing basal area according to our models. Restoration plantings today have goals that go beyond restoring forest cover to an area, ensuring the maintenance of evolutionarily unique native species, and maintaining key ecosystem services and functions. Throughout succession, phylogenetic diversity increased in both treatments without significant differences. We conclude that restoration plans can use eucalyptus as a starting point for restoration plans, especially when carbon sequestration along with biodiversity is the main goal of the restoration project.

Keywords: Atlantic Forest, Biodiversity Conservation, Diversity and Phylogenetic Structure, Ecological restoration, Eucalyptus understory, Forest Plantations, Phylogenetic characters of functional traits, Secondary succession.

INTRODUÇÃO GERAL

A restauração ecológica é o processo de contribuição para a recuperação de um ecossistema que foi degradado, danificado ou destruído (SER, 2004). Além de contribuir para a conservação da biodiversidade, a restauração de ecossistemas visa promover benefícios sociais, econômicos e ambientais, tais como a mitigação das alterações climáticas (CBD, 2019). A Organização das Nações Unidas (ONU) definiu a década 2021-2030 como a Década da Restauração, com o objetivo de deter a degradação dos ecossistemas e restaurá-los para atingir objetivos globais (ONU Brasil, 2021; WRI Brasil, 2021), como o Bonn Challenge (IUCN, 2021), e objetivos nacionais, como o Pacto de Restauração da Mata Atlântica no Brasil, lançado em 2009, que promove a recuperação de 15 milhões de hectares de áreas degradadas até 2050 (PACTO, 2021).

A Mata Atlântica foi identificada pela *Conservation International* como um *hotspot* global devido à fragmentação antrópica e ameaça à biodiversidade (Myers et al., 2000), restando 26,8% da sua cobertura florestal (MapBiomias, 2021). A degradação e o desmatamento na Mata Atlântica foram impulsionados pela expansão das terras agrícolas e monoculturas de árvores (Rodrigues et al., 2009; Rosa et al., 2021), com 57,1% da cobertura em uso extensivo agrícola e pecuário e 3,5% em uso florestal (MapBiomias, 2021). As plantações de eucalipto para fins comerciais no Brasil correspondem a 76,9% das florestas plantadas (IBGE, 2022). Após o abandono do uso da terra, a Mata Atlântica tem um elevado potencial de regeneração natural (Rozendaal et al., 2019; Poorter et al., 2021), quer em antigas áreas de pastagem (Benayas et al., 2007) ou sob plantações de eucalipto (Silva Júnior et al., 1995; Sartori et al., 2002; Souza Filho et al., 2007; Onofre et al., 2010; Viani et al., 2010; Ronquin, 2021).

Para que ocorra a regeneração natural, é necessário o recrutamento de plântulas durante a sucessão natural através do banco de sementes do solo, dispersão de sementes ou rebrotas (Baider et al., 2001; Tabarelli & Peres, 2002; Viani et al., 2015). A montagem de comunidades dirige a sucessão natural através de processos evolutivos e ecológicos baseados na teoria do nicho (concorrência e filtros ambientais) ou teoria neutra (Webb et al., 2002; Cavender-Bares et al., 2009; Meiners et al., 2015). Os filtros ambientais atuam para selecionar espécies dentro de uma comunidade com base na sua capacidade de persistir sob certas condições, limitando a composição de espécies com características ecológicas-funcionais semelhantes (Webb et al., 2002; Cavender-Bares et al., 2009; Kraft & Ackerly, 2010; Bennett et al., 2017). A regeneração natural é eficaz em plantações abandonadas de eucaliptos, que podem catalisar a sucessão florestal no subsolo, mesmo em áreas sem florestas nativas (Viani

et al., 2010; Ronquin, 2021). Por outro lado, o eucalipto pode atuar como um filtro ambiental, influenciando a estrutura filogenética das comunidades através da seleção de espécies semelhantes e estreitamente relacionadas (Kraft et al., 2008; Myers & Harms, 2009).

Duas abordagens revelaram-se promissoras na detecção de respostas na montagem de comunidades às mudanças ambientais: a diversidade filogenética e a funcional. A diversidade filogenética é uma medida que incorpora as relações entre espécies numa comunidade (Magurran, 2004), enquanto que a diversidade funcional incorpora a variação entre espécies e os seus traços que influenciam o modo como as comunidades funcionam (Tilman, 2001). Assim, as medidas que incorporam relações filogenéticas ou traços funcionais podem ser superiores às medidas tradicionais de diversidade (Cianciaruso et al., 2009) e revelaram-se úteis em estudos de restauração florestal (Rother et al., 2019). A diversidade filogenética tem sido utilizada na ecologia de comunidades para inferir a montagem, organização e co-ocorrência de espécies (Webb et al., 2002). Os esforços de restauração visam restaurar um ecossistema com diversidade e função adequadas (Aerts & Honnay, 2011). Isto requer uma compreensão das características funcionais das espécies e pode ajudar a avaliar o sucesso da restauração de acordo com os objetivos e necessidades (Díaz et al., 2013).

Abordamos estas importantes lacunas no conhecimento da diversidade e estrutura filogenética das florestas em sub-bosque de eucalipto na Mata Atlântica, investigando se o eucalipto pode atuar como um filtro ambiental, selecionando espécies filogeneticamente mais relacionadas que outras espécies na comunidade. Como pressuposto, podemos considerar que nas florestas tropicais, a regeneração natural começa com comunidades compostas por poucas espécies pioneiras, com menor riqueza, diversidade e complexidade do que as comunidades clímax encontradas no final da sucessão (Horn, 1974; Magnago et al., 2011; Greenberg et al., 2011; Chazdon, 2012; Crouzeilles et al., 2017; César et al., 2018; Poorter et al., 2021). Assim, esperamos que em áreas abandonadas, o eucalipto possa atuar como um catalisador para a sucessão de espécies nativas no sub-bosque, aumentando a diversidade filogenética à medida que a sucessão avança.

REFERÊNCIAS

Aerts, R., & Honnay, O. (2011). Forest restoration, biodiversity and ecosystem functioning. *BMC Ecology*, 11(29), <https://doi.org/10.1186/1472-6785-11-29>.

Baider, C., Tabarelli, M., & Mantovani, W. The soil seed bank during Atlantic Forest regeneration in Southeast Brazil. *Revista Brasileira de Biologia*, 61(1), 35-44. <https://doi.org/10.1590/S0034-71082001000100006>.

- Bennett, J. A., & Pärtel, M. (2017). Predicting species establishment using absent species and functional neighborhoods. *Ecology and Evolution*, 7(7), 2223-2237. <https://doi.org/10.1002/ece3.2804>.
- Cavender-Bares, J., Kozak, K. H., Fine, P. V. A., & Kembel, S. W. (2009). The merging of community ecology and phylogenetic biology. *Ecology Letters*, 12(7), 693-715. <https://doi.org/10.1111/j.1461-0248.2009.01314.x>.
- CBD. Secretariat and Society for Ecological Restoration. (2019). A companion to the Short-Term Action Plan on Ecosystem Restoration - Resources, cases studies, and biodiversity considerations in the context of restoration science and practice. Montreal, Canada.
- César, R. G., Moreno, V. S., Coletta, G. D., Chazdon, R. L., Ferraz, S. F. B., de Almeida, D. R. A., & Brancalion, P. H. S. (2018). Early ecological outcomes of natural regeneration and tree plantations for restoring agricultural landscapes. *Ecological Applications*, 28(2), 373-384. <https://doi.org/10.1002/eap.1653>.
- Chazdon, R. (2012). Regeneração de florestas tropicais Tropical forest regeneration. *Boletim do Museu Paraense Emílio Goeldi. Ciências Naturais*, 7(3). <https://doi.org/10.46357/bcnaturais.v7i3.587>.
- Cianciaruso, M. V., Silva, I. A., & Antônio Batalha, M. (2009). Diversidades filogenética e funcional: novas abordagens para a Ecologia de comunidades. *Biota Neotrop*, 9(3). <https://doi.org/10.1590/S1676-06032009000300008>.
- Crouzeilles, R., Ferreira, M. S., Chazdon, R. L., Lindenmayer, D. B., B Sansevero, J. B., Monteiro, L., Iribarrem, A., Latawiec, A. E., & N Strassburg, B. B. (2017). Ecological restoration success is higher for natural regeneration than for active restoration in tropical forests. *Science Advances*, 3(11). <https://doi.org/10.1126/sciadv.1701345>.
- Díaz, S., Purvis, A., Cornelissen, J. H. C., Mace, G. M., Donoghue, M. J., Ewers, R. M., Jordano, P., & Pearse, W. D. (2013). Functional traits, the phylogeny of function, and ecosystem service vulnerability. *Ecology and Evolution*, 3(9), 2958-2975. <https://doi.org/10.1002/ece3.601>.
- Greenberg, C. H., Collins, B., Thompson, F. R., & McNab, W. H. (2011). Introduction: What Are Early Successional Habitats, Why Are They Important, and How Can They Be Sustained? In: Greenberg, C., Collins, B., Thompson III, F. (eds) *Sustaining Young Forest Communities. Managing Forest Ecosystems*, Springer, Dordrecht. https://doi.org/10.1007/978-94-007-1620-9_1.
- Horn, H. S. (1974). The ecology of secondary succession. *Annual Review of Ecology and Systematics*, 5, 25-37. <https://doi.org/10.1146/annurev.es.05.110174.000325>.

IBGE. Instituto Brasileiro de Geografia e Estatística. (2022). *Produção da Extração Vegetal e da Silvicultura*. Available in: <https://www.ibge.gov.br/estatisticas/economicas/agricultura-e-pecuaria/9105-producao-da-extracao-vegetal-e-da-silvicultura.html?=&t=sobre>.

IUCN. International Union for Conservation of Nature and Natural Resources. (2021). The Bonn Challenge. Available in: <https://www.iucn.org/theme/forests/our-work/forest-landscape-restoration/bonn-challenge>.

Kraft, N. J. B., & Ackerly, D. D. (2010). Functional trait and phylogenetic tests of community assembly across spatial scales in an Amazonian forest. *Ecological Monographs*, 80(3). <https://doi.org/10.1890/09-1672.1>.

Kraft, N. J. B., Valencia, R., and Ackerly, D. D. (2008). Functional traits and niche-based tree community assembly in an Amazonian forest. *Science* 322(5901), 580-582. <http://dx.doi.org/10.1126/science.1160662>.

Magnago, L. F. S., Simonelli, M., Martins, S. V., Matos, F. A. R., Demuner, V. G. (2011). Variações estruturais e características edáficas em diferentes estádios sucessionais de floresta ciliar de Tabuleiro, ES. *Revista Árvore*, 35(3), 445-456. <https://doi.org/10.1590/S0100-67622011000300008>.

Magurran, A. E. *Measuring biological diversity*. Blackwell, Oxford, p. 256, 2004.

MapBiomas. (2021) Mapeamento Anual da Cobertura e Uso da Terra no Brasil (1985 - 2020) - Destaques Mata Atlântica. Available in: <https://mapbiomas-br-site.s3.amazonaws.com/Fact-Sheet-Mata-Atlantica.pdf>.

Meiners, S. J., Cadotte, M. W., Fridley, J. D., Pickett, S. T. A., & Walker, L. R. (2015). Is successional research nearing its climax? New approaches for understanding dynamic communities. *Functional Ecology*, 29(2), 154-164. <https://doi.org/10.1111/1365-2435.12391>.

Myers, J. A., & Harms, K. E. (2009). Seed arrival, ecological filters, and plant species richness: A meta-analysis. *Ecology Letters*, 12(12), 1250-1260. <https://doi.org/10.1111/j.1461-0248.2009.01373.x>.

Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A. B., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403. <https://doi.org/10.1038/35002501>.

Onofre, F.F., Engel, V.L. & Cassola, H. (2010). Regeneração natural de espécies da Mata Atlântica em sub-bosque de *Eucalyptus saligna* Smith. em uma antiga unidade de produção florestal no Parque das Neblinas, Bertiooga, SP. *Scientia Forestalis*, 38, 39-52.

Poorter, L., Craven, D., Jakovac, C. C., van der Sande, M. T., Amissah, L., Bongers, F., Chazdon, R. L., Farrior, C. E., Kambach, S., Meave, J. A., Muñoz, R., Norden, N., Rüger, N., van Breugel, M.,

Zambrano, A. M. A., Amani, B., Andrade, J. L., Brancalion, P. H. S., Broadbent, E. N., ... Hérault, B. (2021). Multidimensional tropical forest recovery. *Science*, 374(6573), 1370–1376.

<https://doi.org/10.1126/science.abh3629>.

PACTO. Pacto pela Restauração da Mata Atlântica. (2021). Available in:

<https://pactomataatlantica.org.br/o-movimento>.

Rodrigues, R. R., Lima, R. A. F., Gandolfi, S., & Nave, A. G. (2009). On the restoration of high diversity forests: 30 years of experience in the Brazilian Atlantic Forest. *Biological Conservation*, 142(6), 1242-1251. <https://doi.org/10.1016/j.biocon.2008.12.008>.

Ronquin, C. C. (2021). Diversidade de espécies florestais nativas no sub-bosque dos gêneros Eucalyptus e Pinus no Brasil: listagem de 1.136 espécies descritas em 106 trabalhos científicos. Campinas: Embrapa Territorial.

Rosa, M. R., Brancalion, P. H. S., Crouzeilles, R., Tambosi, L. R., Piffer, P. R., Lenti, F. E. B., Hirota, M., Santiami, E., & Metzger, J. P. (2021). Hidden destruction of older forests threatens Brazil's Atlantic Forest and challenges restoration programs. *Science Advances*, 7.

<https://doi.org/10.1126/sciadv.abc4547>.

Rother, D. C., Liboni, A. P., Magnago, L. F. S., Chao, A., Chazdon, R. L., & Rodrigues, R. R. (2019). Ecological restoration increases conservation of taxonomic and functional beta diversity of woody plants in a tropical fragmented landscape. *Forest Ecology and Management*, 451.

<https://doi.org/10.1016/j.foreco.2019.117538>.

Rozendaal, A. D. M., Bongers, F., Mitchell Aide, T., Alvarez-Dávila, E., Ascarrunz, N., Balvanera, P., Becknell, J. M., Bentos, T. v, S Brancalion, P. H., L Cabral, G. A., Calvo-Rodriguez, S., Chave, J., César, R. G., Espírito-Santo, M. M., Fandino, M. C., Wilson Fernandes, G., Finegan, B., García, H., Gonzalez, N., ... Poorter, L. (2019). Biodiversity recovery of Neotropical secondary forests. *Science Advances*, 5(3). <https://doi.org/10.1126/sciadv.aau3114>.

Sartori, M. S., Poggiani, F., Engel, V. L. (2002). Regeneração da vegetação arbórea nativa no sub-bosque de um povoamento de Eucalyptus saligna Smith. Localizado no Estado de São Paulo. *Scientia Forestalis* 62, 59-103.

SER. The SER international primer on ecological restoration. Tucson: Society for Ecological Restoration, 2004.

Silva-Junior, M. C., Scarano, F. R., & Cardel, F. S. (1995). Regeneration of na Atlantic formation in the understory of a Eucalyptus grandis plantation in south-eastern Brazil. *Journal of Tropical Ecology*, 11, 147-152. <https://doi.org/10.1017/S0266467400008518>.

Souza Filho, P.C., Bechara, F.C., Filho, E.M.C., Barreto, K.D., Regeneração natural após diferentes níveis de perturbação em subosque de *Eucalyptus* sp. *Revista Brasileira de Biociências*, 5(1).

Tabarelli, M., & Peres, C. A. (2002). Abiotic and vertebrate seed dispersal in the Brazilian Atlantic forest: implications for forest regeneration. *Biological Conservation*, 106, 165-176.

[https://doi.org/10.1016/S0006-3207\(01\)00243-9](https://doi.org/10.1016/S0006-3207(01)00243-9).

Tilman, D. Functional diversity. In *Encyclopedia of Biodiversity* (S.A. Levin, ed.). Academic Press, San Diego, p. 109-120, 2001.

Viani, R. A. G., Durigan, G., & Melo, A. C. G. de. (2010). A regeneração natural sob plantações florestais: desertos verdes ou redutos de biodiversidade?. *Ciência Florestal*, 20(3).

<https://doi.org/10.5902/198050982067>.

Webb, C. O., Ackerly, D. D., McPeck, M. A., & Donoghue, M. J. (2002). Phylogenies and community ecology. *Annual Review of Ecology and Systematics*, 33, 475-505.

<https://doi.org/10.1146/annurev.ecolsys.33.010802.150448>.

WRI Brasil. Década da Restauração de Ecossistemas é oportunidade para recuperar áreas degradadas no Brasil e no mundo. Available in: <https://wribrasil.org.br/pt/blog/florestas/decada-da-restauracao-ecossistemas-reflorestamento-recuperacao-areas-degradadas-brasil>.

Abandoned Eucalyptus plantations can be used as catalysts for natural regeneration¹

Matheus C. Santos¹, Jeanpierre R. Mirano², Nathália V. H. Safar³, Luiz Fernando S. Magnago^{2*}

¹ *Programa de Pós-Graduação em Ecologia e Conservação da Biodiversidade, Universidade Estadual de Santa Cruz, Ilhéus, Bahia, Brazil;* ² *Centro de Formação em Ciências e Tecnologias Agroflorestais, Universidade Federal do Sul da Bahia, Ilhéus, Bahia, Brazil;* ³ *Departamento de Botânica, Universidade Federal de Viçosa, Viçosa, Minas Gerais, Brazil.*

* Corresponding author. E-mail: m_oak@outlook.com

Acknowledgements: We thank the Coordination for the Improvement of Higher Education Personnel of Brazil (CAPES) for the support to M.C.S (process number 88887.610874/2021-00).

Conflict of Interest: The authors declare no conflict of interest.

Author Contributions: M.C.S. conceived the idea, designed methodology, collected and analyzed the data, and wrote the manuscript. J.R.M. analyzed the data. N.V.H.S. designed methodology and collected the data. L.F.S.M. conceived the idea, designed methodology, collected and analyzed the data, and reviewed the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

Data availability statement: All data used in this manuscript are present in the manuscript and supporting information.

¹ Title page. The manuscript will be submitted to the journal Functional Ecology.

Abstract

1. After abandonment, the Atlantic Forest has a high potential for natural regeneration in areas under eucalyptus plantations. We hypothesize that natural regeneration in tropical forests begins with communities composed of a few pioneer species, which exhibit lower richness, diversity, and complexity than climax communities found at the end of succession. Here, we address diversity and phylogenetic structure in eucalyptus understory forests and investigate whether eucalyptus acts as an environmental filter that selects for more phylogenetically related species.
2. We compared diversity (PD, MPD, MNTD) and phylogenetic structure (sesPD, NRI, NTI) in natural regeneration areas of the Atlantic Forest in two treatments: i) mixed treatment, native species under abandoned eucalyptus plantations; ii) native treatment, with the presence of native species. We established 17 transects in the mixed treatment and 35 in the native treatment. On each transect, we sampled 5-15 contiguous 10 m x 10 m plots. Within each plot, we sampled all live individuals with a diameter at breast height ≥ 10 cm.
3. We found no significant differences between treatments for any of the phylogenetic diversity indices tested. The means of NRI and NTI were positive, indicating phylogenetic clustering in both treatments. None of the phylogenetic diversity and phylogenetic structure metrics were significantly affected by increasing basal area according to our models.
4. Restoration plantings today have goals that go beyond restoring forest cover to an area, ensuring the maintenance of evolutionarily unique native species, and maintaining key ecosystem services and functions. Throughout succession, phylogenetic diversity increased in both treatments without significant differences.
5. We conclude that restoration plans can use eucalyptus as a starting point for restoration plans, especially when carbon sequestration along with biodiversity is the main goal of the restoration project.

Keywords: Atlantic Forest, Biodiversity Conservation, Diversity and Phylogenetic Structure, Ecological restoration, Eucalyptus understory, Forest Plantations, Phylogenetic characters of functional traits, Secondary succession.

Introduction

Ecological restoration is the process of contributing to the recovery of an ecosystem that has been degraded, damaged or destroyed (SER, 2004). In addition to contributing to biodiversity conservation, ecosystem restoration aims to promote social, economic, and environmental benefits, such as climate change mitigation (CBD, 2019). The United Nations (UN) has defined the decade 2021-2030 as the Decade of Restoration, with the aim of halting the degradation of ecosystems and restoring them in order to achieve global goals (UN Brazil, 2021; WRI Brazil, 2021), such as the Bonn Challenge (IUCN, 2021), and national goals, such as the Atlantic Forest Restoration Pact in Brazil, launched in 2009, which promotes the recovery of 15 million hectares of degraded areas by 2050 (PACTO, 2021).

The Atlantic Forest has been identified by Conservation International as a global hotspot for anthropogenic fragmentation and threats to biodiversity (Myers et al., 2000), with 26.8% of its forest cover remaining (MapBiomas, 2021). Degradation and deforestation in the Atlantic Forest have been driven by the expansion of agricultural land and monoculture tree plantations (Rodrigues et al., 2009; Rosa et al., 2021), with 57.1% of the cover in extensive agricultural and livestock use and 3.5% in forestry use (MapBiomas, 2021). *Eucalyptus* plantations for commercial purposes in Brazil correspond to 76.9% of planted forests (IBGE, 2022). After land use abandonment, the Atlantic Forest has a high potential for natural regeneration (Rozendaal et al., 2019; Poorter et al., 2021), either in former pasture areas (Benayas et al., 2007) or under *Eucalyptus* plantations (Silva Júnior et al., 1995; Sartori et al., 2002; Souza Filho et al., 2007; Onofre et al., 2010; Viani et al., 2010; Ronquin, 2021).

For natural regeneration to occur, seedling recruitment is required during natural succession through the soil seed bank, seed dispersal, or stem regrowth (Baider et al., 2001; Tabarelli & Peres, 2002; Viani et al., 2015). Community assembly directs natural succession through evolutionary and ecological processes based on niche theory (competition and environmental filters) or neutral theory (Webb et al., 2002; Cavender-Bares et al., 2009; Meiners et al.,

2015). Environmental filters act to select species within a community based on their ability to persist under certain conditions, limiting the composition of species with similar ecological-functional traits (Webb et al., 2002; Cavender-Bares et al., 2009; Kraft & Ackerly, 2010; Bennett et al., 2017). Natural regeneration is effective in abandoned *Eucalyptus* plantations, which can catalyze forest succession in the understory, even in areas without native forests (Viani et al., 2010; Ronquin, 2021). On the other hand, *Eucalyptus* can act as an environmental filter, influencing the phylogenetic structure of communities by selecting similar and closely related species (Kraft et al., 2008; Myers & Harms, 2009).

Two approaches have shown promise in detecting community responses to environmental change: phylogenetic and functional diversity. Phylogenetic diversity is a measure that incorporates the evolutionary relationships among species in a community (Magurran, 2004), whereas functional diversity incorporates variation among species and their traits that influence how communities function (Tilman, 2001). Thus, measures that incorporate phylogenetic relationships or functional traits may be superior to traditional diversity measures (Cianciaruso et al., 2009) and have proven useful in forest restoration studies (Rother et al., 2019). Phylogenetic diversity has been used in community ecology to infer the assembly, organization, and co-occurrence of community species (Webb et al., 2002). Restoration efforts aim to restore an ecosystem with appropriate diversity and function (Aerts & Honnay, 2011). This requires an understanding of the functional traits of species and can help assess restoration success according to goals and needs (Díaz et al., 2013).

We address these important gaps in our knowledge of the diversity and phylogenetic structure of *Eucalyptus* understory forests in the Atlantic Forest by investigating whether *Eucalyptus* may act as an environmental filter, selecting phylogenetically more closely related species than other species in the community. As an assumption, we can consider that in tropical forests, natural regeneration starts with communities composed of few pioneer species, with

lower richness, diversity and complexity than climax communities found at the end of succession (Horn, 1974; Magnago et al., 2011; Greenberg et al., 2011; Chazdon, 2012; Crouzeilles et al., 2017; César et al., 2018; Poorter et al., 2021). Thus, we expect that in abandoned areas, *Eucalyptus* may act as a catalyst for the succession of native species in the understory, increasing phylogenetic diversity as succession progresses.

Materials and methods

Study area

This study was conducted in the state of Espírito Santo, in the southeast of Brazil. Within the region, we focused on the areas surrounding BR 262, from Viana (km 19) to the district of Vítor Hugo (km 72), and in the basin of the Jucu River (north arm) (Figure 1). The climate is tropical (Aw, Köppen-Geiger classification), with an annual rainfall of 1500 mm and an average annual temperature of 18 °C, ranging from 9.4 °C to 29.6 °C (Feitoza, 1986; Alvares et al., 2013). The vegetation is classified as dense ombrophylous forest and submontane (50-500 m) or montane (500-1500 m), based on the altitude criteria of Garbin et al. (2017). Traditional agricultural use has altered the forests in the region, which have been replaced by forest regeneration or *Eucalyptus* plantations (Simonelli et al., 2021).

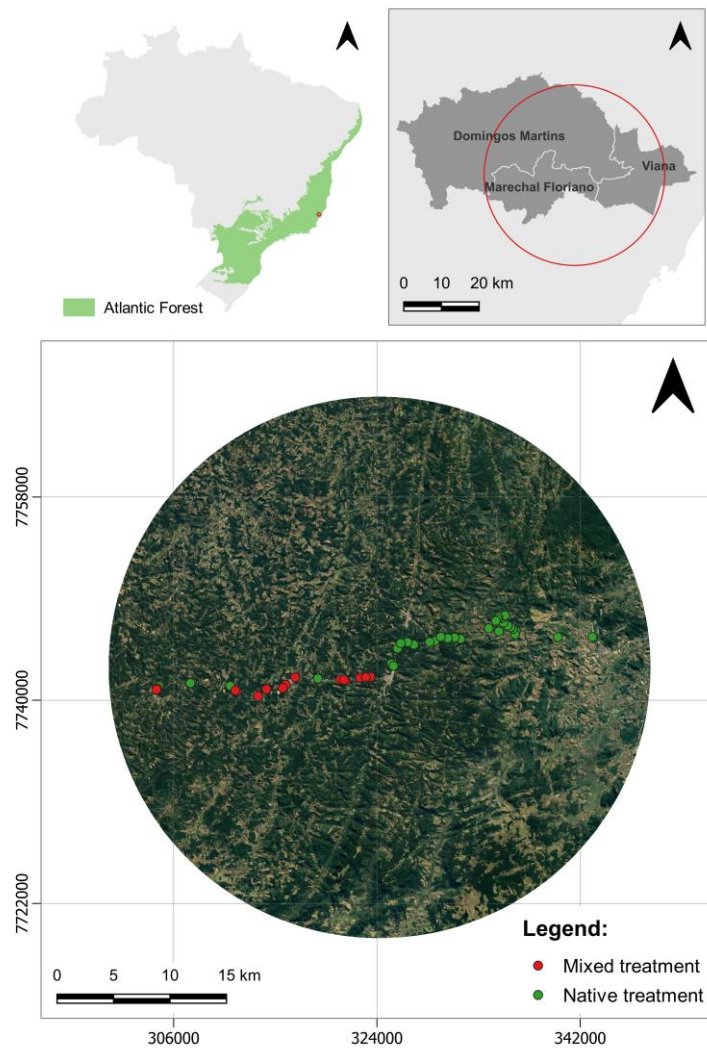


Figure 1. Study sites in the Atlantic Forest of Brazil.

Data collection

We conducted the fieldwork in two natural succession treatments: i) mixed treatment, native species under abandoned *Eucalyptus* plantations; ii) native treatment, with the presence of native species. We sampled 17 transects in the mixed treatment and 35 transects in the native treatment. In each transect, we sampled 5-15 contiguous plots of 10 m x 10 m (0.01 ha), for a total of 494 plots (4.94 ha). Within each plot, we sampled all individuals living and rooted within our plots with a diameter at breast height (DBH; 1.30 m above ground level) ≥ 10 cm. Species not identified in the field were collected, vouchered and subsequently identified through consultations at the Herbarium of Vale (CVRD), Herbarium VIES of the Federal

University of Espírito Santo (UFES) and Professor Mello Leitão Biology Museum (MBML). We classified taxonomy and occurrence in Brazilian biomes based on the Flora of Brazil database (BFG, 2021); We checked spellings, taxonomic status and establishment in PlantMiner (Carvalho et al., 2010).

Measures of phylogenetic diversity and structure

To construct our phylogeny, we generated a list of all our species, genera, and families based on the Angiosperm Phylogeny Group IV (APG, 2016). We omitted *Araucaria* Juss. and *Pinus* L. (Gymnosperms) because of their very distinct and ancient phylogenetic lineages. The list elaborated with the selected species was added to the GBOTB.extended.tre phylogenetic tree using the V.PhyloMaker package (Jin & Qian, 2019) in R, version 4.2.2 (R Core Team, 2022).

To explain phylogeny, we use the following metrics (Faith, 1992; Webb et al, 2002; Swenson, 2014): (i) Phylogenetic Diversity (PD), found by summing phylogenetic branches for all species and expressed in millions of years; (ii) Mean Pairwise Distance (MPD), mean phylogenetic distance between all combinations of pairs of individuals; (iii) Mean Nearest Taxon Distance (MNTD), mean phylogenetic distance between an individual and the closest related (nonspecific) individual. In addition, we calculated the standardized effect size (ses) of PD, Net Relatedness Index (NRI), and Nearest Taxon Index (NTI) to eliminate the wealth effect based on a null model. Negative values indicate phylogenetic clustering and positive values indicate phylogenetic overdispersion. We calculated these six metrics using the Picante package (Kembel et al., 2010) in R, version 4.2.2 (R Core Team, 2022). For standard effect size calculations, we compared our tree to 999 random null models using the taxa.labels algorithm, which incorporates the labels from the community data matrix, and without considering the relative abundance of species in the communities (abundance.weighted = FALSE).

Functional trait matrix

We examined four traits (Table 1) related to morphological and physical adaptations of trees in their role as dispersers, food resources, forest structure and carbon storage: i) dispersal syndrome according to Van der Pijl (1982); ii) seed size according to Tabarelli & Peres (2002); iii) maximum height; iv) wood density. We classified the maximum height and wood density traits as follows: small/low ($1 < (\mu - \sigma)$); medium ($(\mu - \sigma) \leq 2 < (\mu + \sigma)$); large/high ($3 \geq (\mu + \sigma)$), where μ = mean and σ = sample standard deviation.

Table 1. Functional traits used to calculate the functional diversity value.

Functional trait	Relevance	Unit	Categories
Dispersal syndrome	Colonization capacity	cat ¹	1. zoochoric 2. non-zoochoric
Seed size	Quantity and type of food resource	cm	1. small (< 0.6) 2. medium ($0.6-1.6$) 3. large ($\geq 1.6-3.0$) 4. extremely large (> 3.0)
Maximum height	Competitive potential and growth period	m	1. small (< 9.64) 2. medium ($9.64-29.78$) 3. large (≥ 29.78)
Wood density	Potential for carbon allocation in biomass	g cm ⁻³	1. low (< 0.438) 2. medium ($0.438-0.708$) 3. high (≥ 0.708)

¹ cat = categorical.

Data for dispersal syndrome, seed size, and maximum height were obtained from the TRY database, the REFLORA program, and the literature, in addition to collecting field material. We obtained data for wood density from the Global Wood Density Database (Chave et al., 2009; Zanne et al., 2009) through the BIOMASS package (Réjou-Méchain et al., 2017) in R, version 4.2.2 (R Core Team, 2022), in the Tropical South America subsection. The list of data sources used in the study can be found in the Data sources section.

Phylogenetic signal of the functional traits

To quantify the phylogenetic signal in the functional trait data, we used the K statistic (Blomberg et al., 2003), which compares the observed signal to another under a Brownian motion model of trait evolution in a phylogeny. In this model, $K = 0$ indicates a lack of phylogenetic signal; $K > 1$ indicates that lineages are more similar than expected, causing clustering; $K < 1$ indicates that lineages are more divergent than expected, indicating overdispersal. The statistical significance of the phylogenetic signal was evaluated by comparing the observed patterns of trait-independent contrast variance with a null model of taxon label shuffling at the top of the phylogeny. We computed K-statistics and randomizations with the multiPhylosignal function using the Picante package (Kembel et al., 2010) in R, version 4.2.2 (R Core Team, 2022).

Data analysis

To assess how phylogenetic diversity differed between treatments, we compared the means of the phylogenetic metric indices (PD, MPD, MNTD, sesPD, NRI, NTI) in the mixed and native treatments using Student's t-test with a significance level less than or equal to 0.05 ($P \leq 0.05$) through the stats package in R, version 4.2.2 (R Core Team, 2022). We tested the normality of the residuals using the Shapiro-Wilk test with the RVAideMemoire package in R, version 4.2.2 (R Core Team, 2022).

To assess how phylogenetic diversity affects succession, we used basal area of individual trees as a proxy for succession ($BA = \frac{\pi DBH^2}{40000}$; Soares et al., 2006) and phylogenetic indices as response variables. We then constructed generalized linear models with Gaussian error distributions, implemented in the glm function of the statistics package in R, version 4.2.2 (R Core Team, 2022). We also considered possible interactions between the predictor variables (treatments*basal area). The regression models were visualized using the visreg package in R,

version 4.2.2 (R Core Team, 2022).

Results

In total, we recorded 3240 individuals of 215 tree species from 140 genera and 51 families according to APG IV (2016). The mean and standard deviation of species richness for the mixed treatment was 16.05 ± 5.78 (range: 6-29 species), and 17.40 ± 7.42 (range: 4-37 species) for the native treatment. The most representative species in the both treatments were *Moquiniastrum polymorphum* (291), *Croton floribundus* (254), and *Piptadenia gonoacantha* (206). The most representative families in the both treatments were Fabaceae, Asteraceae, and Euphorbiaceae. The phylogenetic tree was constructed with 215 tips and 183 internal nodes (Figure S1). The mean and standard deviation of basal area of native species was $23.75 \text{ m}^2/\text{ha} \pm 13.89 \text{ m}^2/\text{ha}$ (range: 2.87-54.74 m^2/ha) in the mixed treatment, and $24.29 \text{ m}^2/\text{ha} \pm 8.83 \text{ m}^2/\text{ha}$ (range: 6.58-48.09 m^2/ha) in the native treatment. We verified the average time to abandonment of the *Eucalyptus* plantations based on the mean DBH, which ranged from 20.05 cm to 52.01 cm, with a mean of $37.22 \text{ cm} \pm 8.52 \text{ cm}$. The mean DBH value for *Eucalyptus* plantations in the state of Espírito Santo is 35.6 cm after 21 years of planting (Nutto et al., 2006). Therefore, we can estimate that the *Eucalyptus* species have at least 20 years of abandonment.

Does Eucalyptus act as an environmental filter?

We found that PD (Pearson: $r = 0.969$, $P < 0.05$) and MNTD (Pearson: $r = -0.592$, $P < 0.05$) were significantly correlated with species richness. We found no significant correlation between SR and MPD (Pearson: $r = 0.244$, $P = 0.08$), sesPD (Pearson: $r = 0.088$, $P = 0.53$), NRI (Pearson: $r = -0.248$, $P = 0.07$) and NTI (Pearson: $r = 0.016$, $P = 0.9066$). Thus, standardized values minimized the effects of species richness (Figure S2).

We found no significant differences between treatments for any of the phylogenetic diversity

metrics tested (Table S1). The mean PD was 0.11% higher in the mixed treatment than in the native treatment, harboring phylogenetic diversity with a species composition spanning 1536.02 million years (Fig. 2a). The mean MPD was 0.33% lower in the mixed treatment than in the native treatment, with more related species and a mean distance of 235.10 million years (Fig. 2b). The mean MNTD was 3.17% smaller in the mixed treatment than in the native treatment, with a species composition with a mean nearest neighbor distance of 135.98 million years, indicating that the terminal species in this treatment are more closely related (Fig. 2c).

We found no significant differences between treatments for any of the phylogenetic structure indices tested (Table S1). The means of sesPD were negative, indicating a lower PD than expected by chance (Fig. 2d). The means of NRI and NTI were positive, indicating phylogenetic clustering in both treatments (Figs. 2e-2f).

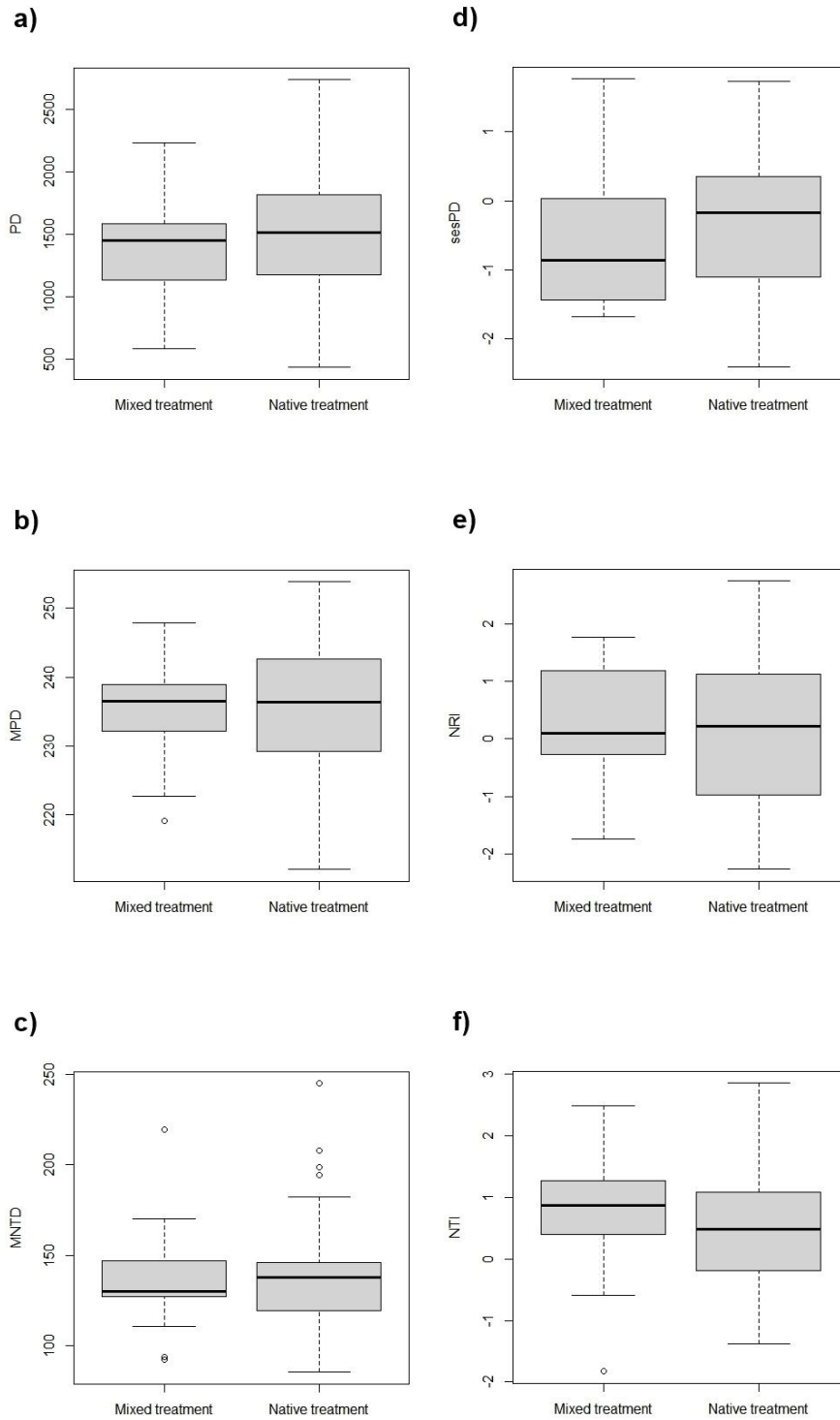


Figure 2. Patterns of a) Phylogenetic Diversity (PD), b) Mean Peer Distance (MPD), c) Mean Nearest Taxon Distance (MNTD), d) Standardized Effect Size (ses) of PD, e) Net Relatedness Index (NRI), and f) Nearest Taxon Index (NTI), in mixed and native treatments of natural regeneration in the Atlantic Forest.

Does phylogenetic diversity increase over succession?

None of the phylogenetic diversity (i.e., PD; MPD and MNTD) and phylogenetic structure (i.e., sesPD, NRI and NTI) metrics were significantly affected by increasing basal area according to our models. The interactions between the predictor variables were also not significant.

For phylogenetic diversity, our models showed that PD was positively affected by increasing basal area in the mixed (slope: 6.380) and native (slope: 20.974) treatments (Fig. 3a). MPD was positively affected by increasing basal area in the mixed (slope: 0.013) and native (slope: 0.299) treatments (Fig. 3b). The MNTD data did not show normality of residuals, so we removed outliers older than 200 million years (Figure S3). MNTD increased with increasing basal area in the mixed and native treatments (Fig. 3c).

For phylogenetic structure, our models showed that sesPD was positively affected by increasing basal area in the mixed (slope: 0.018) and native (slope: 0.033) treatments (Fig. 3d). PD was correlated with species richness (Figure S2). When the effect of richness was eliminated and the observed PD values were compared with those of communities generated by the null models (Table S2), communities with higher PD are expected to have positive sesPD values and communities with lower PD are expected to have negative sesPD values (Figs. 3a-3d).

In the mixed treatment, NRI was negatively affected and NTI was positively affected with increasing basal area (Figs. 3e-3f). However, the trend line remained positive for both NRI and NTI, indicating more phylogenetically clustered species than expected by chance.

In the native treatment, NRI and NTI were negatively affected with increasing basal area (Figs. 3e-3f). When the effect of richness was removed (Figs. 3b-3c), NRI and NTI tended to be negative throughout succession, indicating phylogenetic overdispersion with less closely related species than expected by chance.

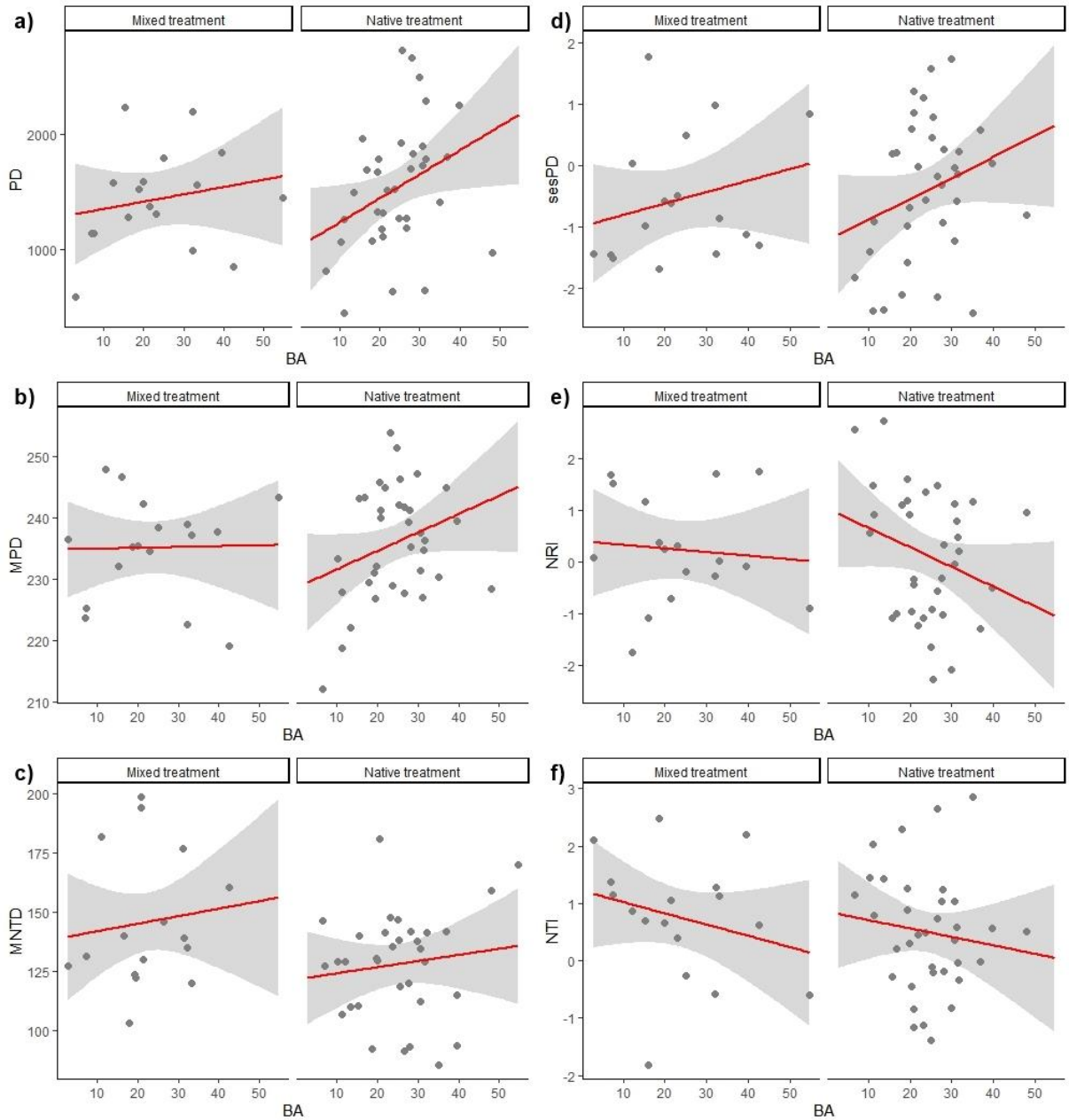


Figure 3. Plots of models of basal area of native species (BA) in relation to diversity indices and phylogenetic structure: a) Phylogenetic Diversity (PD), b) Mean Peer Distance (MPD), c) Mean Nearest Taxon Distance (MNTD), d) Standardized Effect Size (ses) of PD, e) Net Relatedness Index (NRI), and f) Nearest Taxon Index (NTI), in mixed and native treatments of natural regeneration in the Atlantic Forest. The red line represents the fitted curve and the gray areas are the 95% confidence intervals.

Are functional traits conserved across lineages?

We found a phylogenetic signal for all traits in the mixed and native treatments, with K values less than 1, indicating that the traits are more phylogenetically clustered than expected by chance (Table 2). The P-value was less than 0.05 for most functional traits (except for maximum tree height), indicating that most of the traits analyzed were less variable than expected from the random phylogeny of the community (Table 2). Given that most traits had significant phylogenetic sign, close relatives generally have more similar trait values than would be expected by chance.

Table 2. Results of phylogenetic signal analysis in functional traits using Blomberg's K.

Functional Traits	Mixed treatment		Native treatment	
	K	P	K	P
Dispersal syndrome	0.537	0.001	0.709	0.001
Seed size	0.292	0.001	0.213	0.002
Maximum height	0.189	0.057	0.205	0.002
Wood density	0.561	0.001	0.399	0.001

K = Blomberg's K index; P = significance level (≤ 0.05).

Discussion

The potential of abandoned *Eucalyptus* plantations as a catalyst for natural regeneration has been evaluated in several studies, but it is not known how these communities are structured and what the phylogenetic diversity of the *Eucalyptus* understory is (Viani et al., 2010; Ronquin, 2021). Knowledge of the evolutionary history of species and their functions in ecosystems helps to guide restoration strategies (Aerts & Honnay, 2011). Currently, restoration plantings have goals beyond restoring forest cover in an area, such as ensuring the maintenance of evolutionarily unique native species (Rodrigues et al., 2009; Charles, 2018) and maintaining key ecosystem services and functions, such as carbon stocks (Pan et al., 2011; Magnago et al., 2015; Koch & Kaplan, 2022). Here, we show that for most metrics tested, phylogenetic diversity does not change in areas where natural regeneration has

occurred in abandoned *Eucalyptus* plantations compared to areas that regenerated under other conditions (pasture or abandoned coffee plantations). During succession, phylogenetic diversity increased in both treatments, with closely related species sharing similar traits. Our results therefore suggest that restoration plans can use *Eucalyptus* as a starting point for restoration plans, especially when carbon sequestration is the main goal of the restoration project along with biodiversity (Brancalion et al., 2018; Brancalion et al., 2020).

Does Eucalyptus act as an environmental filter?

Our results showed that there was no significant difference between the means of the phylogenetic diversity indices in the treatments, with similar PD, MPD and MNTD values in the mixed and native treatments. Some studies have shown that the negative effects of *Eucalyptus* on the regeneration of native species, suggesting that for species with certain life history traits, the *Eucalyptus* understory may be ineffective for regeneration, acting as an environmental filter (Durigan et al., 2004; Ostertag et al., 2008; Pairo et al., 2021). In other biomes, such as the Cerrado, natural regeneration under *Eucalyptus* plantations has had poorer results due to cover and biomass interference (see Durigan et al., 2004). In ecosystems in other countries, the *Eucalyptus* understorey was dominated by exotic species from Hawaii due to light conditions (see Ostertag et al., 2008). There are several filters acting on the development of understory regeneration, both in *Eucalyptus* plantations and in native forests. In planted monocultures, there are additional filters to those already acting in native forests, such as: historical and environmental factors, canopy density, light availability in the understory, age of planting, planted forest species, distance to remnants of native vegetation, management of planted forests, and history of use of the area, which can directly or indirectly influence the richness, density, and structure of natural regeneration under *Eucalyptus* plantations (Geldenhuys, 1997; Parrotta, 1999; Viani et al., 2010).

Our results showed that there was no significant difference between the means of the

phylogenetic structure indices in the treatments, with similar sesPD, NRI and NTI values in the mixed and native treatments. The observed PD values are similar to those obtained for the simulated communities with the redistribution of species names in the phylogeny, with only 13% of the communities showing a $P < 0.05$ value (Table S2). On average, NTI and NRI indicate the occurrence of phylogenetic clustering in both treatments, with ecologically similar and more phylogenetically related species. This means that the species represent communities with an analogous amount of evolutionary history. A possible explanation, is due to the proximity of regenerating areas with matricial forests (250 meters) in the Central Mountainous Zone of Espírito Santo, with about 80.26% of its area presenting potential to receive propagules (Simonelli et al., 2021). The distance of the area to be restored from forest fragments providing propagules is strategic for the success of natural regeneration (Chazdon, 2012; Simonelli et al., 2021). *Eucalyptus* plantations can increase landscape connectivity, i.e., the dispersal and movement of native species. In many tropical regions, animals are the main seed dispersers and are therefore important for the dynamics of plant regeneration (Parrotta et al., 1997; Lindenmayer & Hobbs, 2004; Coelho et al., 2022).

Does phylogenetic diversity increase with succession?

We found no significant differences in diversity indices and phylogenetic structure as succession progressed in the treatments. In both treatments, we observed that phylogenetic diversity remained constant throughout the succession. The use of *Eucalyptus* as a facilitator species for the restoration of native forest vegetation has shown positive results, as they can act as pioneer species and trigger the initial successional process (Viani et al., 2010; Ronquin, 2021). This technique of intercropping *Eucalyptus* species with native species is an effective tool for forest restoration (Brancalion et al., 2018; Brancalion et al., 2020), especially in degraded areas where native species would not perform well due to major barriers to growth, such as drought, fire, and functional degradation of soil fertility and structure (Campello,

1998; Magnago et al., 2011; Ronquin, 2021). Our results confirm that competition with native trees does not appear to be strong enough to affect their survival or to suppress native trees during succession, which could have reduced species diversity and their evolutionary history (see Brancalion et al., 2020). These results may provide a new strategy for initiating forest restoration plans, as Brazilian native tree species grow more slowly than species of the genus *Eucalyptus* (Ronquin, 2021), although competition for resources with *Eucalyptus* can slow the growth of native species (Durigan et al., 2004; Brancalion et al., 2020; Pairo et al., 2021). Thus, in addition to the arrival of invertebrate and vertebrate species (da Rocha et al., 2012; Chazdon, 2012; Martin et al., 2012; Timo et al., 2015; Jacoboski et al., 2016) that provide important seed dispersal services in the early stages of restoration, abandoned *Eucalyptus* areas provide important habitat for native species (Viani et al., 2010; Ronquin, 2021; Coelho et al., 2022).

In the native treatment, we observed increased phylogenetic diversity during succession. We also observed that as succession progresses, species increase their relatedness by introducing species that are phylogenetically more distant throughout the succession. Thus, in early successional stages, communities are clustered by the effect of the environmental filter, whereas in advanced successional stages they exhibit phylogenetic dispersal due to limiting similarity (Letcher et al., 2012), resulting in the competitive exclusion of ecologically similar and phylogenetically close species (Webb et al., 2002; Cavender-Bares et al., 2009). We did not find this trend in *Eucalyptus* plantations due to the age of the regenerating community, which recruits pioneer species in younger plantations and subsequently starts to favor the establishment of species later in the ecological succession (Carneiro & Rodrigues, 2007; Pairo et al., 2021). The most abundant pioneer species were *Croton floribundus*, *Myrcia splendens*, *Piptadenia gonoacantha*, and *Senna multijuga* (see Carvalho, 2014; Lorenzi, 2021).

Are functional traits conserved across lineages?

Because most of our functional traits had significant phylogenetic signal, close relatives generally share more similar trait values than would be expected by chance. We quantified the phylogenetic signal of four functional traits for the species in the mixed and native treatments, which describes how selective pressures and life history act on the phenotypic evolution of species (Blomberg et al., 2003). All K values (Table 2) for the phylogenetic signal were less than one, suggesting that the traits are more labile than expected under a Brownian model of trait evolution. Thus, environmental filters would select functionally similar species based on their ability to adapt to the same environmental conditions, bringing together species with similar ecological niches (Cavender-Bares et al., 2009). The results for the phylogenetic signal of the traits analyzed indicate that wood density and dispersal syndrome are phylogenetically less convergent than expected by the Brownian model, but phylogenetically more conserved than a random association between traits and phylogeny. In addition, related species tend to be more similar than random in wood density and dispersal syndrome in the mixed treatment, and dispersal syndrome in the native treatment.

Dispersal syndrome is a trait associated with the establishment and distribution of species in new areas, and is related to dispersal distance and community structure (Cornelissen et al., 2003). Of the species found in our study area, 74.88% have animal dispersal (Data sources). Some studies cite the predominance of zoochoric species among regenerating individuals (Viani et al., 2010; Callegaro et al., 2013; Simonelli et al., 2021; Ronquin, 2021; Coelho et al., 2022), dispersal with greater influence in tropical forests due to the complexity of the environment and the co-evolutionary relationships between species (Silva et al., 1996). On the other hand, seed arrivals of non-zoochoric species tend to decrease as eucalypts grow because they become an obstacle to wind flow (Keenan et al., 1997).

Wood density follows successional trends: early in succession, pioneer species with acquisitive strategies and low wood density; throughout succession, slow-growing species

with conservative strategies and high wood density (Chazdon, 2012; Poorter et al., 2019). Younger *Eucalyptus* plantations are expected to have greater recruitment of pioneer species, which gradually decreases with increasing shading, favoring the establishment of species later in the ecological succession. (Carneiro & Rodrigues, 2007). This may explain the high representation of pioneer species in the mixed treatment, as they have similar characteristics and are functionally similar to *Eucalyptus*.

References

- Aerts, R., & Honnay, O. (2011). Forest restoration, biodiversity and ecosystem functioning. *BMC Ecology*, *11*(29), <https://doi.org/10.1186/1472-6785-11-29>.
- Alvares, C. A., Stape, J. L., Sentelhas, P. C., de Moraes, J. L. G., & Sparovek, G. (2013). Köppen's climate classification map for Brazil. *Meteorologische Zeitschrift*, *22*(6), 711-728. <https://doi.org/10.1127/0941-2948/2013/0507>.
- APG (The Angiosperm Phylogeny Group). (2016). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants. *Botanical Journal of the Linnean Society*, *181*(1), 1-20. <https://doi.org/10.1111/boj.12385>.
- Baider, C., Tabarelli, M., & Mantovani, W. The soil seed bank during Atlantic Forest regeneration in Southeast Brazil. *Revista Brasileira de Biologia*, *61*(1), 35-44. <https://doi.org/10.1590/S0034-71082001000100006>.
- Benayas, J. M. R., Martins, A., Nicolau, J. M., & Schulz, J. J. (2007). Abandonment of agricultural land: An overview of drivers and consequences. *CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources* *57*(2), 1-14. <https://doi.org/10.1079/PAVSNNR20072057>.
- Bennett, J. A., & Pärtel, M. (2017). Predicting species establishment using absent species and functional neighborhoods. *Ecology and Evolution*, *7*(7), 2223-2237. <https://doi.org/10.1002/ece3.2804>.
- BFG (The Brazil Flora Group). (2021). Coleção Flora do Brasil 2020. Data from: Jardim Botânico do Rio de Janeiro. <http://doi.org/10.47871/jbrj2021004>.
- Blomberg, S. P., Garland, T., & Ives, A. R. (2003). Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution*, *57*(4), 717-745, <https://doi.org/10.1111/j.0014-3820.2003.tb00285.x>.

Brancalion, P. H. S., Bello, C., Chazdon, R. L., Galetti, M., Jordano, P., Lima, R. A. F., Medina, A., Pizo, M. A., & Reid, J. L. (2018). Maximizing biodiversity conservation and carbon stocking in restored tropical forests. *Conservation Letters*, *11*(4). <https://doi.org/10.1111/conl.12454>.

Brancalion, P. H. S., Amazonas, N. T., Chazdon, R. L., van Melis, J., Rodrigues, R. R., Silva, C. C., Sorrini, T. B., & Holl, K. D. (2020). Exotic eucalypts: From demonized trees to allies of tropical forest restoration? *Journal of Applied Ecology*, *57*(1), 55-66. <https://doi.org/10.1111/1365-2664.13513>.

Callegaro, R. M., Andrzejewski, C., Longhi, S. J., Araujo, M. M., & Serra, G. C. (2013). Potencial de três plantações florestais homogêneas como facilitadoras da regeneração natural de espécies arbutivo-arbóreas. *Scientia Forestalis*, *41*(99), 331-341.

Campello, E. F. C. (1998). Sucessão vegetal na recuperação de áreas degradadas, In DIAS, L.E., & MELLO, J.W.V. (Eds.) – Recuperação de áreas degradadas, Viçosa: UFV, Departamento de Solos; Sociedade Brasileira de Recuperação de Áreas Degradadas, p. 183-202.

Carneiro, P. H. M., Rodrigues, R. R. (2007). Management of monospecific commercial reforestations for the forest restoration of native species with high diversity. In: RODRIGUES, R. R. et al. High Diversity Forest Restoration in Degraded Areas: Methods and Projects in Brazil. New York: Nova Science Publishers, 129-144.

Carvalho, G. H., Cianciaruso, M. V., & Batalha, M. A. (2010). Plantminer: A web tool for checking and gathering plant species taxonomic information. *Environmental Modelling and Software*, *25*(6), 815-816. <https://doi.org/10.1016/j.envsoft.2009.11.014>.

Carvalho, P. E. R. (2014). Espécies Arbóreas Brasileiras. Colombo: Embrapa Florestas.

Cavender-Bares, J., Kozak, K. H., Fine, P. V. A., & Kembel, S. W. (2009). The merging of community ecology and phylogenetic biology. *Ecology Letters*, *12*(7), 693-715. <https://doi.org/10.1111/j.1461-0248.2009.01314.x>.

CBD. Secretariat and Society for Ecological Restoration. (2019). A companion to the Short-Term Action Plan on Ecosystem Restoration - Resources, cases studies, and biodiversity considerations in the context of restoration science and practice. Montreal, Canada.

César, R. G., Moreno, V. S., Coletta, G. D., Chazdon, R. L., Ferraz, S. F. B., de Almeida, D. R. A., & Brancalion, P. H. S. (2018). Early ecological outcomes of natural regeneration and tree plantations for restoring agricultural landscapes. *Ecological Applications*, *28*(2), 373-384. <https://doi.org/10.1002/eap.1653>.

- Charles, L. S. (2018). Plant Functional Traits and Species Selection in Tropical Forest Restoration. *Tropical Conservation Science*, 11. <https://doi.org/10.1177/1940082918784157>.
- Chave, J., Coomes, D., Jansen, S., Lewis, S. L., Swenson, N. G., & Zanne, A. E. (2009). Towards a worldwide wood economics spectrum. *Ecology Letters*, 12(4), 351-366. <https://doi.org/10.1111/j.1461-0248.2009.01285.x>.
- Chazdon, R. (2012). Regeneração de florestas tropicais Tropical forest regeneration. *Boletim do Museu Paraense Emílio Goeldi. Ciências Naturais*, 7(3). <https://doi.org/10.46357/bcnaturais.v7i3.587>.
- Cianciaruso, M. V., Silva, I. A., & Antônio Batalha, M. (2009). Diversidades filogenética e funcional: novas abordagens para a Ecologia de comunidades. *Biota Neotrop*, 9(3). <https://doi.org/10.1590/S1676-06032009000300008>.
- Coelho, A. J. P., Villa, P. M., Matos, F. A. R., Heringer, G., Bueno, M. L., de Paula Almado, R., & Meira-Neto, J. A. A. (2022). Atlantic Forest recovery after long-term eucalyptus plantations: The role of zoochoric and shade-tolerant tree species on carbon stock. *Forest Ecology and Management*, 503. <https://doi.org/10.1016/j.foreco.2021.119789>.
- Cornelissen, J. H. C., Lavorel, S., Garnier, E., Díaz, S., Buchmann, N., Gurvich, D. E., Reich, P. B., ter Steege, H., Morgan, H. D., van der Heijden, M. G. A., Pausas, J. G., & Poorter, H. (2003). A handbook of protocols for standardised and easy measurement of plant functional traits worldwide. *Australian Journal of Botany*, 51(4), 335-380. <https://doi.org/10.1071/BT02124>.
- Crouzeilles, R., Ferreira, M. S., Chazdon, R. L., Lindenmayer, D. B., B Sansevero, J. B., Monteiro, L., Iribarrem, A., Latawiec, A. E., & N Strassburg, B. B. (2017). Ecological restoration success is higher for natural regeneration than for active restoration in tropical forests. *Science Advances*, 3(11). <https://doi.org/10.1126/sciadv.1701345>.
- Díaz, S., Purvis, A., Cornelissen, J. H. C., Mace, G. M., Donoghue, M. J., Ewers, R. M., Jordano, P., & Pearse, W. D. (2013). Functional traits, the phylogeny of function, and ecosystem service vulnerability. *Ecology and Evolution*, 3(9), 2958-2975. <https://doi.org/10.1002/ece3.601>.
- Durigan, G., Carlos, A., Melo, G. de, Contieri, W. A., & Nakata, H. (2004). Regeneração Natural da Vegetação de Cerrado sob Florestas Plantadas com Espécies Nativas e Exóticas. In: VILAS BÔAS, O.; DURIGAN, G. Pesquisas em conservação e recuperação ambiental no Oeste Paulista: resultados da cooperação Brasil/Japão. São Paulo: Páginas e Letras, 2004. p. 349-362.
- Faith, D. P. (1992). Conservation evaluation and phylogenetic diversity. *Biological Conservation*, 61(1), 1-10. [https://doi.org/10.1016/0006-3207\(92\)91201-3](https://doi.org/10.1016/0006-3207(92)91201-3).

- Feitoza, L. R. (1986). *Carta agroclimática do Espírito Santo*. Data from: Empresa Capixaba de Pesquisa Agropecuária. Available in: <https://biblioteca.incaper.es.gov.br/digital/bitstream/123456789/2986/1/carta-agroclimatica-do-ES.jpg>.
- Garbin, M. L., Saiter, F. Z., Carrijo, T. T., & Peixoto, A. L. (2017). Breve histórico e classificação da vegetação capixaba. *Rodriguesia*, 68(5), 1883-1894. <https://doi.org/10.1590/2175-7860201768521>.
- Geldenhuys, C. J. (1997). Native forest regeneration in pine and eucalypt plantations in Northern Province, South Africa. *Forestry Ecology and Management*, 99, 101-115. [https://doi.org/10.1016/S0378-1127\(97\)00197-7](https://doi.org/10.1016/S0378-1127(97)00197-7).
- Greenberg, C. H., Collins, B., Thompson, F. R., & McNab, W. H. (2011). Introduction: What Are Early Successional Habitats, Why Are They Important, and How Can They Be Sustained? In: Greenberg, C., Collins, B., Thompson III, F. (eds) *Sustaining Young Forest Communities. Managing Forest Ecosystems*, Springer, Dordrecht. https://doi.org/10.1007/978-94-007-1620-9_1.
- Horn, H. S. (1974). The ecology of secondary succession. *Annual Review of Ecology and Systematics*, 5, 25-37. <https://doi.org/10.1146/annurev.es.05.110174.000325>.
- IBGE. Instituto Brasileiro de Geografia e Estatística. (2022). *Produção da Extração Vegetal e da Silvicultura*. Available in: <https://www.ibge.gov.br/estatisticas/economicas/agricultura-e-pecuaria/9105-producao-da-extracao-vegetal-e-da-silvicultura.html?=&t=sobre>.
- IUCN. International Union for Conservation of Nature and Natural Resources. (2021). The Bonn Challenge. Available in: <https://www.iucn.org/theme/forests/our-work/forest-landscape-restoration/bonn-challenge>.
- Jakovac, C. C., Junqueira, A. B., Crouzeilles, R., Peña-Claros, M., Mesquita, R. C. G., & Bongers, F. (2021). The role of land-use history in driving successional pathways and its implications for the restoration of tropical forests. *Biological Reviews*, 96(4), 1114-1134. <https://doi.org/10.1111/brv.12694>.
- Jacoboski, L. I., Debastiani, V. J., de Mendonça-Lima, A., & Hartz, S. M. (2016). How do diversity and functional nestedness of bird communities respond to changes in the landscape caused by eucalyptus plantations? *Community Ecology Community Ecology*, 17(1), 107-113. <https://doi.org/10.1556/168.2016.17.1.13>.
- Jin, Y., & Qian, H. (2019). V.PhyloMaker: an R package that can generate very large phylogenies for vascular plants. *Ecography*, 42(8), 1353-1359. <https://doi.org/10.1111/ecog.04434>.

Keenan, R., Lamb, D., Woldring, O., Irvine, T., & Jensen, R. (1997). Restoration of plant biodiversity beneath tropical tree plantations in Northern Australia. *Forest Ecology and Management*, 99, 117-131. [https://doi.org/10.1016/S0378-1127\(97\)00198-9](https://doi.org/10.1016/S0378-1127(97)00198-9).

Kembel, S. W., Cowan, P. D., Helmus, M. R., Cornwell, W. K., Morlon, H., Ackerly, D. D., Blomberg, S. P., & Webb, C. O. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26(11), 1463-1464. <https://doi.org/10.1093/bioinformatics/btq166>.

Koch, A., & Kaplan, J. O. (2022). Tropical forest restoration under future climate change. *Nature Climate Change*, 12(3), 279-283. <https://doi.org/10.1038/s41558-022-01289-6>.

Kraft, N. J. B., & Ackerly, D. D. (2010). Functional trait and phylogenetic tests of community assembly across spatial scales in an Amazonian forest. *Ecological Monographs*, 80(3). <https://doi.org/10.1890/09-1672.1>.

Kraft, N. J. B., Valencia, R., and Ackerly, D. D. (2008). Functional traits and niche-based tree community assembly in an Amazonian forest. *Science* 322(5901), 580-582. <http://dx.doi.org/10.1126/science.1160662>.

Letcher, S. G., Chazdon, R. L., Andrade, A. C. S., Bongers, F., van Breugel, M., Finegan, B., Laurance, S. G., Mesquita, R. C. G., Martínez-Ramos, M., & Williamson, G. B. (2012). Phylogenetic community structure during succession: Evidence from three Neotropical forest sites. *Perspectives in Plant Ecology, Evolution and Systematics*, 14(2), 79-87. <https://doi.org/10.1016/j.ppees.2011.09.005>.

Lindenmayer, D. B., & Hobbs, R. J. (2004). Fauna conservation in Australian plantation forests - A review. *Biological Conservation*, 119(2), 151-168. <https://doi.org/10.1016/j.biocon.2003.10.028>.

Lorenzi, H. Árvores Brasileiras: Manual de Identificação e Cultivo de Plantas Arbóreas do Brasil. Nova Odessa, SP: Instituto Plantarum, 3ª edição, 2021.

Magnago, L. F. S., Simonelli, M., Martins, S. V., Matos, F. A. R., Demuner, V. G. (2011). Variações estruturais e características edáficas em diferentes estádios sucessionais de floresta ciliar de Tabuleiro, ES. *Revista Árvore*, 35(3), 445-456. <https://doi.org/10.1590/S0100-67622011000300008>.

Magnago, L. F. S., Magrath, A., Laurance, W. F., Martins, S. v., Meira-Neto, J. A. A., Simonelli, M., & Edwards, D. P. (2015). Would protecting tropical forest fragments provide carbon and biodiversity cobenefits under REDD+? *Global Change Biology*, 21(9), 3455-3468. <https://doi.org/10.1111/gcb.12937>.

Magurran, A. E. Measuring biological diversity. Blackwell, Oxford, p. 256, 2004.

- MapBiomass. (2021) Mapeamento Anual da Cobertura e Uso da Terra no Brasil (1985 - 2020) - Destaques Mata Atlântica. Available in: <https://mapbiomas-br-site.s3.amazonaws.com/Fact-Sheet-Mata-Atlantica.pdf>.
- Martin, P. S., Gheler-Costa, C., Lopes, P. C., Rosalino, L. M., & Verdade, L. M. (2012). Terrestrial non-volant small mammals in agro-silvicultural landscapes of Southeastern Brazil. *Forest Ecology and Management*, 282, 185-195. <https://doi.org/10.1016/j.foreco.2012.07.002>.
- Meiners, S. J., Cadotte, M. W., Fridley, J. D., Pickett, S. T. A., & Walker, L. R. (2015). Is successional research nearing its climax? New approaches for understanding dynamic communities. *Functional Ecology*, 29(2), 154-164. <https://doi.org/10.1111/1365-2435.12391>.
- Myers, J. A., & Harms, K. E. (2009). Seed arrival, ecological filters, and plant species richness: A meta-analysis. *Ecology Letters*, 12(12), 1250-1260. <https://doi.org/10.1111/j.1461-0248.2009.01373.x>.
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A. B., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403. <https://doi.org/10.1038/35002501>.
- Nutto, L., Spathelf, P., Seling, I., & Florestal, E. (2006). Management of individual tree diameter growth and implications for pruning for Brazilian Eucalyptus grandis Hill ex Maiden. *Floresta*, 36(3). <http://dx.doi.org/10.5380/RF.v36i3.7519>.
- Onofre, F.F., Engel, V.L. & Cassola, H. (2010). Regeneração natural de espécies da Mata Atlântica em sub-bosque de Eucalyptus saligna Smith. em uma antiga unidade de produção florestal no Parque das Neblinas, Bertioga, SP. *Scientia Forestalis*, 38, 39-52.
- Ostertag, R., Giardina, C. P., & Cordell, S. (2008). Understory colonization of eucalyptus plantations in Hawaii in relation to light and nutrient levels. *Restoration Ecology*, 16(3), 475-485. <https://doi.org/10.1111/j.1526-100X.2007.00321.x>.
- PACTO. Pacto pela Restauração da Mata Atlântica. (2021). Available in: <https://pactomataatlantica.org.br/o-movimento>.
- Pairo, P. E., Rodriguez, E. E., Bellocq, M. I., & Aceñolaza, P. G. (2021). Changes in taxonomic and functional diversity of plants in a chronosequence of Eucalyptus grandis plantations. *Scientific Reports*, 11(1). <https://doi.org/10.1038/s41598-021-89988-6>.
- Pan, Y., Birdsey, R. A., Jingyun F., Houghton, R., Kauppi, P. E., Kurz, W. A., Phillips, O. L., Shvidenko, A., Lewis, S. L., Canadell, J. G., Ciais, P., Jackson, R. B., Pacala, S. W., McGuire, A. D., Piao, S., Rautiainen, A., Sitch, S., Hayes, D. (2011). A Large and Persistent Carbon Sink in the World's Forests. *Science*, 333(6045), 988-993. <https://doi.org/10.1126/science.1201609>.

Parrotta, J. A., Turnbull, J. W., & Jones, N. (1997). Introduction Catalyzing native forest regeneration on degraded tropical lands. *Forestry Ecology and Management*, 99, 1-7.

[https://doi.org/10.1016/S0378-1127\(97\)00190-4](https://doi.org/10.1016/S0378-1127(97)00190-4).

Parrotta, J. A. (1999). Productivity, nutrient cycling, and succession in single- and mixed-species plantations of *Casuarina equisetifolia*, *Eucalyptus robusta*, and *Leucaena leucocephala* in Puerto Rico. *Forestry Ecology and Management*, 124(1), 45-77. [https://doi.org/10.1016/S0378-1127\(99\)00049-3](https://doi.org/10.1016/S0378-1127(99)00049-3).

Poorter, L., Rozendaal, D. M. A., Bongers, F., de Almeida-Cortez, J. S., Almeyda Zambrano, A. M., Álvarez, F. S., Andrade, J. L., Villa, L. F. A., Balvanera, P., Becknell, J. M., Bentos, T. v., Bhaskar, R., Boukili, V., Brancalion, P. H. S., Broadbent, E. N., César, R. G., Chave, J., Chazdon, R. L., Colletta, G. D., ... Westoby, M. (2019). Wet and dry tropical forests show opposite successional pathways in wood density but converge over time. *Nature Ecology and Evolution*, 3(6), 928-934.

<https://doi.org/10.1038/s41559-019-0882-6>.

Poorter, L., Craven, D., Jakovac, C. C., van der Sande, M. T., Amissah, L., Bongers, F., Chazdon, R. L., Fariior, C. E., Kambach, S., Meave, J. A., Muñoz, R., Norden, N., Rüger, N., van Breugel, M., Zambrano, A. M. A., Amani, B., Andrade, J. L., Brancalion, P. H. S., Broadbent, E. N., ... Hérault, B. (2021). Multidimensional tropical forest recovery. *Science*, 374(6573), 1370–1376.

<https://doi.org/10.1126/science.abh3629>.

R Core Team. (2022). R: A language and environment for statistical computing. Data from: R Foundation for Statistical Computing. Available in: <https://www.R-project.org/>.

Réjou-Méchain, M., Tanguy, A., Piponiot, C., Chave, J., & Hérault, B. (2017). biomass: an r package for estimating above-ground biomass and its uncertainty in tropical forests. *Methods in Ecology and Evolution*, 8(9), 1163-1167. <https://doi.org/10.1111/2041-210X.12753>.

da Rocha, P. L. B., Viana, B. F., Cardoso, M. Z., de Melo, A. M. C., Costa, M. G. C., de Vasconcelos, R. N., & Dantas, T. B. (2013). What is the value of eucalyptus monocultures for the biodiversity of the Atlantic forest? A multitaxa study in southern Bahia, Brazil. *Journal of Forestry Research*, 24(2), 263-272. <https://doi.org/10.1007/s11676-012-0311-z>.

Rodrigues, R. R., Lima, R. A. F., Gandolfi, S., & Nave, A. G. (2009). On the restoration of high diversity forests: 30 years of experience in the Brazilian Atlantic Forest. *Biological Conservation*, 142(6), 1242-1251. <https://doi.org/10.1016/j.biocon.2008.12.008>.

Ronquin, C. C. (2021). Diversidade de espécies florestais nativas no sub-bosque dos gêneros *Eucalyptus* e *Pinus* no Brasil: listagem de 1.136 espécies descritas em 106 trabalhos científicos. Campinas: Embrapa Territorial.

Rother, D. C., Liboni, A. P., Magnago, L. F. S., Chao, A., Chazdon, R. L., & Rodrigues, R. R. (2019). Ecological restoration increases conservation of taxonomic and functional beta diversity of woody plants in a tropical fragmented landscape. *Forest Ecology and Management*, 451.

<https://doi.org/10.1016/j.foreco.2019.117538>.

Rosa, M. R., Brancalion, P. H. S., Crouzeilles, R., Tambosi, L. R., Piffer, P. R., Lenti, F. E. B., Hirota, M., Santiami, E., & Metzger, J. P. (2021). Hidden destruction of older forests threatens Brazil's Atlantic Forest and challenges restoration programs. *Science Advances*, 7.

<https://doi.org/10.1126/sciadv.abc4547>.

Rozendaal, A. D. M., Bongers, F., Mitchell Aide, T., Alvarez-Dávila, E., Ascarrunz, N., Balvanera, P., Becknell, J. M., Bentos, T. v, S Brancalion, P. H., L Cabral, G. A., Calvo-Rodriguez, S., Chave, J., César, R. G., Espírito-Santo, M. M., Fandino, M. C., Wilson Fernandes, G., Finegan, B., García, H., Gonzalez, N., ... Poorter, L. (2019). Biodiversity recovery of Neotropical secondary forests. *Science Advances*, 5(3). <https://doi.org/10.1126/sciadv.aau3114>.

Sartori, M. S., Poggiani, F., Engel, V. L. (2002). Regeneração da vegetação arbórea nativa no sub-bosque de um povoamento de Eucalyptus saligna Smith. Localizado no Estado de São Paulo. *Scientia Forestalis* 62, 59-103.

SER. The SER international primer on ecological restoration. Tucson: Society for Ecological Restoration, 2004.

Silva, J. M. C., Uhl, C., & Murray, G. (1996). Society for Conservation Biology Plant Succession, Landscape Management, and the Ecology of Frugivorous Birds in Abandoned Amazonian Pastures. *Biology* 10(2).

Silva-Junior, M. C., Scarano, F. R., & Cardel, F. S. (1995). Regeneration of na Atlantic formation in the understory of a Eucalyptus grandis plantation in south-eastern Brazil. *Journal of Tropical Ecology*, 11, 147-152. <https://doi.org/10.1017/S0266467400008518>.

Simonelli, M., Martins, S. V., Sartori, M., Raposo Filho, F. L., Dadalto, G., & Pereira, M. L. (2021). Levantamento do potencial de regeneração natural de florestas nativas nas diferentes regiões do estado do Espírito Santo (1st ed.). Edifes.

Soares, C. P. B., Paula Neto, F., Souza, A. L. (2006). Dendrometria e Inventário Florestal. Viçosa: UFV.

Souza Filho, P.C., Bechara, F.C., Filho, E.M.C., Barreto, K.D., Regeneração natural após diferentes níveis de perturbação em subosque de Eucalyptus sp. *Revista Brasileira de Biociências*, 5(1).

Swenson, N. G. (2014). *Use R! Functional and Phylogenetic Ecology in R* (1st ed.). Springer.
<https://doi.org/10.1007/978-1-4614-9542-0>.

Tabarelli, M., & Peres, C. A. (2002). Abiotic and vertebrate seed dispersal in the Brazilian Atlantic forest: implications for forest regeneration. *Biological Conservation*, *106*, 165-176.
[https://doi.org/10.1016/S0006-3207\(01\)00243-9](https://doi.org/10.1016/S0006-3207(01)00243-9).

Tilman, D. Functional diversity. In *Encyclopedia of Biodiversity* (S.A. Levin, ed.). Academic Press, San Diego, p. 109-120, 2001.

Timo, T. P. C., Lyra-Jorge, M. C., Gheler-Costa, C., & Verdade, L. M. (2014). Effect of the plantation age on the use of eucalyptus stands by medium to large-sized wild mammals in south-eastern Brazil. *IForest*, *8*, 108-113. <https://doi.org/10.3832/IFOR1237-008>.

UN Brazil. (2021). *Começa a Década da ONU da Restauração de Ecossistemas*. Available in:
<https://brasil.un.org/pt-br/130341-comeca-decada-da-onu-da-restauracao-de-ecossistemas>.

Van der Pijl, L. (1982). *Principles of Dispersal in Higher Plants* (3rd ed.). Springer Verlag.

Viani, R. A. G., Durigan, G., & Melo, A. C. G. de. (2010). A regeneração natural sob plantações florestais: desertos verdes ou redutos de biodiversidade?. *Ciência Florestal*, *20*(3).
<https://doi.org/10.5902/198050982067>.

Webb, C. O., Ackerly, D. D., McPeck, M. A., & Donoghue, M. J. (2002). Phylogenies and community ecology. *Annual Review of Ecology and Systematics*, *33*, 475-505.
<https://doi.org/10.1146/annurev.ecolsys.33.010802.150448>.

WRI Brasil. *Década da Restauração de Ecossistemas é oportunidade para recuperar áreas degradadas no Brasil e no mundo*. Available in: <https://wribrasil.org.br/pt/blog/florestas/decada-da-restauracao-ecossistemas-reflorestamento-recuperacao-areas-degradadas-brasil>.

Zanne, A. E., Lopez-Gonzalez, G., Coomes, D. A., Ilic, J., Jansen, S., Lewis, S. L., Miller, R. B., Swenson, N. G., Wiemann, M. C., & Chave, J. (2009) Data from: Towards a worldwide wood economics spectrum. Dryad, Dataset. <https://doi.org/10.5061/dryad.234>.

Data sources

N	Specie	Family	meanWD	levelWD	Maximum height	Data	Dispersal syndrome	Data	Seed size	Data
1	Abarema jupunba	Fabaceae	0.5851	genus	35	Try	zoo	Try	3.5	Bonadeu, 2013
2	Aegiphila integrifolia	Lamiaceae	0.6450	genus	14.25	Try	zoo	Try	0.1	REFLORA
3	Albizia polycephala	Fabaceae	0.5317	genus	17	Safar et al., 2022	zoo	Try	0.68	Safar et al., 2022
4	Alchornea glandulosa	Euphorbiaceae	0.4013	genus	25.65	Try	zoo	Try	0.44	Safar et al., 2022
5	Alchornea triplinervia	Euphorbiaceae	0.4013	genus	27.59	Try	zoo	Try	0.5	Santos & Caruzo, 2014
6	Allagoptera caudescens	Arecaceae	0.5604	family	12.71	Safar et al., 2022	zoo	Safar et al., 2022	0.18	Safar et al., 2022
7	Allophylus edulis	Sapindaceae	0.5604	genus	19	Try	zoo	Try	0.57	Safar et al., 2022
8	Allophylus petiolulatus	Sapindaceae	0.4350	genus	12	Safar et al., 2022	zoo	Safar et al., 2022	0.62	Safar et al., 2022
9	Allophylus puberulus	Sapindaceae	0.4350	genus	20	REFLORA	zoo	Try	0.5	Sommer, 2009
10	Allophylus racemosus	Sapindaceae	0.4350	genus	9	BDC	zoo	Try	1	REFLORA
11	Alseis floribunda	Rubiaceae	0.7500	genus	14.97	Safar et al., 2022	nzoo	Try	0.1	Safar et al., 2022
12	Amaioua	Rubiaceae	0.6250	genus	7	Try	zoo	Try	0.5	REFLORA
13	Amaioua intermedia	Rubiaceae	0.6250	genus	13.08	Safar et al., 2022	zoo	Try	0.96	Safar et al., 2022
14	Anadenanthera	Fabaceae	0.8113	genus	19.49	Safar et al., 2022	nzoo	Safar et al., 2022	0.1	Safar et al., 2022
15	Andira legalis	Fabaceae	0.7543	genus	14.65	Safar et al., 2022	zoo	Safar et al., 2022	0.45	Safar et al., 2022
16	Annona acutirlora	Annonaceae	0.4240	genus	7	Safar et al., 2022	zoo	Safar et al., 2022	0.51	Safar et al., 2022
17	Annona cacans	Annonaceae	0.4240	genus	14.33	Safar et al., 2022	zoo	Safar et al., 2022	0.63	Safar et al., 2022
18	Annona dolabripetala	Annonaceae	0.4240	genus	19.29	Safar et al., 2022	zoo	Safar et al., 2022	0.45	Safar et al., 2022
19	Aparisthium cordatum	Euphorbiaceae	0.3900	genus	7.88	Safar et al., 2022	nzoo	Safar et al., 2022	0.33	Safar et al., 2022
20	Apuleia leiocarpa	Fabaceae	0.7690	genus	15	Try	nzoo	Safar et al., 2022	0.62	Safar et al., 2022
21	Artocarpus heterophyllus	Moraceae	0.5167	genus	20	Try	nzoo	Try	3.26	Silva et al., 2010
22	Astrocaryum aculeatissimum	Arecaceae	0.5080	genus	14.75	Try	zoo	Try	3.04	Safar et al., 2022
23	Astronium graveolens	Anacardiaceae	0.8254	genus	39.52	Try	zoo	Try	0.18	Safar et al., 2022
24	Bathysa australis	Rubiaceae	0.5604	family	26.6	Try	nzoo	Try	0.1	Junior & Vieira, 2015
25	Bauhinia longifolia	Fabaceae	0.6000	genus	15	Safar et al., 2022	nzoo	Try	0.92	Safar et al., 2022
26	Brosimum glaucum	Moraceae	0.6577	genus	25.63	Safar et al., 2022	zoo	Safar et al., 2022	1.06	Safar et al., 2022
27	Byrsonima	Malpighiaceae	0.6287	genus	15.5	Safar et al., 2022	zoo	Try	0.78	Safar et al., 2022
28	Byrsonima sericea	Malpighiaceae	0.6287	genus	18.5	Safar et al., 2022	zoo	Try	0.5	Safar et al., 2022
29	Byrsonima stipulacea	Malpighiaceae	0.6287	genus	15.5	Safar et al., 2022	zoo	Safar et al., 2022	1.02	Safar et al., 2022
30	Cabralea canjerana	Meliaceae	0.4775	genus	40	Try	zoo	Try	2	REFLORA
31	Cariniana estrellensis	Lecythidaceae	0.5476	genus	11.82	Safar et al., 2022	nzoo	Try	0.72	Safar et al., 2022
32	Cariniana legalis	Lecythidaceae	0.5476	genus	35	Safar et al., 2022	nzoo	Safar et al., 2022	0.82	Safar et al., 2022
33	Caryota urens	Arecaceae	0.5604	family	10	REFLORA	zoo	Try	0.15	Pimenta, 2015
34	Casearia arborea	Salicaceae	0.6777	genus	32.48	Try	zoo	Try	0.2	Marquete & Vaz, 2007
35	Casearia commersoniana	Salicaceae	0.6777	genus	12.7	Try	zoo	Try	0.6	Safar et al., 2022
36	Casearia decandra	Salicaceae	0.6777	genus	20	Try	zoo	Safar et al., 2022	0.7	Marquete & Vaz, 2007
37	Casearia espiritosantensis	Salicaceae	0.6777	genus	6	BDC	zoo	Try	0.2	REFLORA
38	Casearia javitensis	Salicaceae	0.6777	genus	18	Try	zoo	Try	0.4	Marquete & Zappi, 2018
39	Casearia sylvestris	Salicaceae	0.6777	genus	21.85	Try	zoo	Try	0.25	Marquete & Vaz, 2007
40	Casearia ulmifolia	Salicaceae	0.6777	genus	20	Try	zoo	Try	0.15	Safar et al., 2022
41	Cathedra rubricaulis	Aptandraceae	0.5604	SouthAmerica	8	Lucena et al., 2020	zoo	Lucena et al., 2020	0.2	Lucena et al., 2020
42	Cecropia glaziovii	Urticaceae	0.3077	genus	23.75	Try	zoo	Try	0.15	FFESP
43	Cecropia hololeuca	Urticaceae	0.3077	genus	20.71	Safar et al., 2022	zoo	Try	0.18	FFESP
44	Cecropia pachystachya	Urticaceae	0.3077	genus	15.06	Safar et al., 2022	zoo	Try	0.61	Safar et al., 2022

45	<i>Cedrela fissilis</i>	Meliaceae	0.3988	genus	30	Try	zoo	Safar et al., 2022	4	REFLORA
46	<i>Chrysophyllum</i>	Sapotaceae	0.7677	genus	25	Safar et al., 2022	zoo	Safar et al., 2022	2.2	Safar et al., 2022
47	<i>Citronella</i>	Cardioperidaceae	0.5604	family	11	BDC	zoo	Safar et al., 2022	1.15	Safar et al., 2022
48	<i>Clarisia ilicifolia</i>	Moraceae	0.5302	genus	18	Safar et al., 2022	zoo	Try	1.09	Safar et al., 2022
49	<i>Clitoria fairchildiana</i>	Fabaceae	0.5604	family	12	Lorenzi, 2021	nzoo	Try	0.16	Embrapa
50	<i>Clusia spiritu-sanctensis</i>	Clusiaceae	0.6000	genus	8	FFESP	zoo	FFESP	0.15	FFESP
51	<i>Copaifera langsdorffii</i>	Fabaceae	0.6081	genus	22.33	Safar et al., 2022	zoo	Try	1.18	Safar et al., 2022
52	<i>Cordia magnoliifolia</i>	Boraginaceae	0.4874	genus	7.5	Safar et al., 2022	zoo	Safar et al., 2022	0.66	Safar et al., 2022
53	<i>Cordia trichotoma</i>	Boraginaceae	0.4874	genus	11.33	Safar et al., 2022	zoo	Safar et al., 2022	1.1	Safar et al., 2022
54	<i>Couepia</i>	Chrysobalanaceae	0.8046	genus	25.59	Safar et al., 2022	zoo	Safar et al., 2022	2.2	Safar et al., 2022
55	<i>Coussapoa microcarpa</i>	Urticaceae	0.4623	genus	24.68	Try	zoo	Try	1.2	Safar et al., 2022
56	<i>Coussarea contracta</i>	Rubiaceae	0.6103	genus	19.95	Try	zoo	Try	2.24	Safar et al., 2022
57	<i>Croton floribundus</i>	Euphorbiaceae	0.4071	genus	10.86	Safar et al., 2022	nzoo	Safar et al., 2022	0.33	Safar et al., 2022
58	<i>Cupania</i>	Sapindaceae	0.6193	genus	16	BDC	zoo	Safar et al., 2022	2	REFLORA
59	<i>Cupania oblongifolia</i>	Sapindaceae	0.6193	genus	6.5	Safar et al., 2022	zoo	Try	1.5	FFESP
60	<i>Cupania racemosa</i>	Sapindaceae	0.6193	genus	13.92	Safar et al., 2022	zoo	Try	2	FFESP
61	<i>Cupania rugosa</i>	Sapindaceae	0.6193	genus	18.36	Safar et al., 2022	zoo	Safar et al., 2022	2	REFLORA
62	<i>Cupania scrobiculata</i>	Sapindaceae	0.6193	genus	22	Try	zoo	Try	1.1	Safar et al., 2022
63	<i>Cupania vernalis</i>	Sapindaceae	0.6193	genus	23.75	Try	zoo	Try	1.7	FFESP
64	<i>Cupania zanthoxyloides</i>	Sapindaceae	0.6193	genus	15	REFLORA	zoo	Try	2.7	FFESP
65	<i>Dalbergia nigra</i>	Fabaceae	0.8084	genus	20	Try	nzoo	Safar et al., 2022	0.48	Safar et al., 2022
66	<i>Dialium guianense</i>	Fabaceae	0.8673	genus	40	Try	zoo	Try	0.56	Safar et al., 2022
67	<i>Dictyoloma vandellianum</i>	Rutaceae	0.5604	family	13.77	Safar et al., 2022	nzoo	Embrapa	0.21	Safar et al., 2022
68	<i>Didymopanax calvus</i>	Araliaceae	0.5750	genus	30	REFLORA	zoo	Barbosa et al., 2016	0.6	FFESP
69	<i>Endlicheria paniculata</i>	Lauraceae	0.5011	genus	21.85	Try	zoo	Try	2.5	FFESP
70	<i>Eriotheca candolleana</i>	Malvaceae	0.4407	genus	18.42	Safar et al., 2022	nzoo	Try	0.73	Safar et al., 2022
71	<i>Eriotheca macrophylla</i>	Malvaceae	0.4407	genus	30.34	Safar et al., 2022	nzoo	Safar et al., 2022	0.57	Safar et al., 2022
72	<i>Erythroxylum pulchrum</i>	Erythroxylaceae	0.7100	genus	5.5	Safar et al., 2022	zoo	Safar et al., 2022	0.65	Safar et al., 2022
73	<i>Eschweilera ovata</i>	Lecythidaceae	0.8253	genus	27.97	Safar et al., 2022	zoo	Safar et al., 2022	1.77	Safar et al., 2022
74	<i>Eucaliptus</i>	Myrtaceae	0.5604	family	40	REFLORA	nzoo	REFLORA	0.25	REFLORA
75	<i>Eugenia</i>	Myrtaceae	0.7219	genus	25	Safar et al., 2022	zoo	Safar et al., 2022	1.81	Safar et al., 2022
76	<i>Eugenia zuccarinii</i>	Myrtaceae	0.7219	genus	25	Safar et al., 2022	zoo	Safar et al., 2022	2	REFLORA
77	<i>Euphorbiaceae</i>	Euphorbiaceae	0.5604	family	6.79	Safar et al., 2022	zoo	Safar et al., 2022	3.27	Safar et al., 2022
78	<i>Euterpe edulis</i>	Arecaceae	0.4065	genus	19	Try	zoo	Try	1.1	Safar et al., 2022
79	<i>Fabaceae</i>	Fabaceae	0.5604	family	7.54	Safar et al., 2022	nzoo	Safar et al., 2022	1.18	Safar et al., 2022
80	<i>Ficus</i>	Moraceae	0.3956	genus	40	Try	zoo	Try	0.1	Safar et al., 2022
81	<i>Ficus clusiifolia</i>	Moraceae	0.3956	genus	40	Try	zoo	Try	0.05	REFLORA
82	<i>Ficus insipida</i>	Moraceae	0.3956	genus	40	Try	zoo	Try	0.05	REFLORA
83	<i>Gallesia integrifolia</i>	Phytolaccaceae	0.4800	genus	30	Try	nzoo	Try	3	REFLORA
84	<i>Garcinia gardneriana</i>	Clusiaceae	0.6567	genus	8.5	Safar et al., 2022	zoo	Safar et al., 2022	1.48	Safar et al., 2022
85	<i>Geissospermum laeve</i>	Apocynaceae	0.7823	genus	31	Try	zoo	Try	1.1	Safar et al., 2022
86	<i>Guapira</i>	Nyctaginaceae	0.4923	genus	25	Safar et al., 2022	zoo	Try	0.63	Safar et al., 2022
87	<i>Guapira noxia</i>	Nyctaginaceae	0.4923	genus	9	Safar et al., 2022	zoo	Try	0.2	Safar et al., 2022
88	<i>Guapira opposita</i>	Nyctaginaceae	0.4923	genus	25	Safar et al., 2022	zoo	Try	0.63	Safar et al., 2022
89	<i>Guarea guidonia</i>	Meliaceae	0.6319	genus	40	Try	zoo	Try	1	REFLORA
90	<i>Guatteria</i>	Annonaceae	0.5554	genus	12.23	Safar et al., 2022	zoo	Safar et al., 2022	0.4	Safar et al., 2022
91	<i>Guatteria sellowiana</i>	Annonaceae	0.5554	genus	36.08	Safar et al., 2022	zoo	Safar et al., 2022	0.65	Safar et al., 2022
92	<i>Guettarda viburnoides</i>	Rubiaceae	0.7067	genus	8.49	Safar et al., 2022	zoo	Safar et al., 2022	1.33	Safar et al., 2022

93	<i>Himatanthus bracteatus</i>	Apocynaceae	0.5021	genus	13.83	Safar et al., 2022	nzoo	Safar et al., 2022	0.98	Safar et al., 2022
94	<i>Hirtella insignis</i>	Chrysobalanaceae	0.7955	genus	7	Safar et al., 2022	zoo	Safar et al., 2022	0.8	Safar et al., 2022
95	<i>Hyeronima alchorneoides</i>	Phyllanthaceae	0.5604	family	40	Try	zoo	Try	0.5	Lorenzi, 2021
96	<i>Inga capitata</i>	Fabaceae	0.5813	genus	30	Try	zoo	Safar et al., 2022	0.73	Safar et al., 2022
97	<i>Inga laurina</i>	Fabaceae	0.5813	genus	30	Try	zoo	Try	1	REFLORA
98	<i>Inga marginata</i>	Fabaceae	0.5813	genus	30	Try	zoo	Try	1	REFLORA
99	<i>Inga subnuda</i>	Fabaceae	0.5813	genus	14.56	Safar et al., 2022	zoo	Try	1	Safar et al., 2022
100	<i>Inga vera</i>	Fabaceae	0.5813	genus	50	Try	zoo	Try	1.5	REFLORA
101	<i>Jacaranda puberula</i>	Bignoniaceae	0.3954	genus	23.49	Try	nzoo	Safar et al., 2022	0.677	Safar et al., 2022
102	<i>Jacaratia heptaphylla</i>	Caricaceae	0.2650	genus	17.68	Safar et al., 2022	zoo	Safar et al., 2022	0.3	Safar et al., 2022
103	<i>Jacaratia spinosa</i>	Caricaceae	0.2650	genus	40	Try	zoo	Try	0.7	REFLORA
104	<i>Kielmeyera</i>	Calophyllaceae	0.5604	family	8	Ramos, 2011	zoo	Try	1	Ramos, 2011
105	<i>Ladenbergia hexandra</i>	Rubiaceae	0.4900	genus	11	FFESP	nzoo	FFESP	0.11	FFESP
106	<i>Lamanonia ternata</i>	Cunoniaceae	0.5604	family	19	Try	nzoo	Try	0.05	Oliveira et al., 2019
107	Lauraceae	Lauraceae	0.5604	family	22.04	Safar et al., 2022	zoo	Safar et al., 2022	2.09	Safar et al., 2022
108	<i>Licania kunthiana</i>	Chrysobalanaceae	0.8287	genus	45	Try	zoo	Try	1.98	Safar et al., 2022
109	<i>Licaria guianensis</i>	Lauraceae	0.7726	genus	19	Try	zoo	Safar et al., 2022	0.2	Safar et al., 2022
110	<i>Lonchocarpus cultratus</i>	Fabaceae	0.7073	genus	19.9	Safar et al., 2022	nzoo	Safar et al., 2022	0.57	Safar et al., 2022
111	<i>Machaerium aculeatum</i>	Fabaceae	0.4947	genus	12	Lorenzi, 2021	nzoo	Try	1.5	Lorenzi, 2021
112	<i>Machaerium hirtum</i>	Fabaceae	0.4947	genus	12	IPÊ	nzoo	Try	1.4	Martins et al., 2016
113	<i>Machaerium nyctitans</i>	Fabaceae	0.4947	genus	18	Lorenzi, 2021	nzoo	Try	1.4	Filho et al., 2006
114	<i>Mangifera indica</i>	Anacardiaceae	0.5604	family	30	Try	zoo	Try	8.42	Coral & Garcia, 2021
115	<i>Matayba guianensis</i>	Sapindaceae	0.7713	genus	24	Try	zoo	Try	0.82	Safar et al., 2022
116	Maytenus	Celastraceae	0.7446	genus	6	Barbosa et al., 2016	zoo	Barbosa et al., 2016	4.5	REFLORA
117	<i>Miconia</i>	Melastomataceae	0.6229	genus	14.05	Safar et al., 2022	zoo	Safar et al., 2022	0.6	Safar et al., 2022
118	<i>Miconia calvescens</i>	Melastomataceae	0.6229	genus	8	Try	zoo	Try	0.75	Chagas, 2012
119	<i>Miconia cinnamomifolia</i>	Melastomataceae	0.6229	genus	24.7	Try	zoo	Safar et al., 2022	0.6	Safar et al., 2022
120	<i>Miconia holosericea</i>	Melastomataceae	0.6229	genus	7.75	Try	zoo	Try	0.2	Chagas, 2012
121	<i>Miconia prasina</i>	Melastomataceae	0.6229	genus	15.25	Try	zoo	Try	0.63	Safar et al., 2022
122	<i>Micropholis</i>	Sapotaceae	0.6568	genus	33.41	Safar et al., 2022	zoo	Safar et al., 2022	0.8	Safar et al., 2022
123	<i>Monteverdia obtusifolia</i>	Celastraceae	0.7446	genus	8.68	Safar et al., 2022	zoo	Safar et al., 2022	0.5	Safar et al., 2022
124	<i>Moquilea tomentosa</i>	Chrysobalanaceae	0.5604	family	20	REFLORA	zoo	Coradin et al., 2018	6	Coradin et al., 2018
125	<i>Moquiniastrum polymorphum</i>	Asteraceae	0.5604	SouthAmerica	10	REFLORA	nzoo	Faria, 2016	0.3	Faria, 2016
126	<i>Myrcia neoglabra</i>	Myrtaceae	0.8068	genus	13	Safar et al., 2022	zoo	Safar et al., 2022	1	Safar et al., 2022
127	<i>Myrcia splendens</i>	Myrtaceae	0.8068	genus	30	Try	zoo	Try	0.46	Safar et al., 2022
128	<i>Myrcia vittoriana</i>	Myrtaceae	0.8068	genus	9.62	Safar et al., 2022	zoo	Try	0.35	Safar et al., 2022
129	<i>Myrciaria tenella</i>	Myrtaceae	0.6575	genus	6	REFLORA	zoo	Try	0.4	REFLORA
130	<i>Myrsine coriacea</i>	Primulaceae	0.5604	family	21.85	Try	zoo	Try	0.24	Safar et al., 2022
131	<i>Myrsine guianensis</i>	Primulaceae	0.5604	family	11.23	Safar et al., 2022	zoo	Try	0.26	Safar et al., 2022
132	Myrtaceae	Myrtaceae	0.5604	family	16.5	Safar et al., 2022	zoo	Safar et al., 2022	0.48	Safar et al., 2022
133	<i>Nectandra membranacea</i>	Lauraceae	0.5207	genus	35	Try	zoo	Try	0.1	Goldenberg & Moraes, 2009
134	<i>Nectandra oppositifolia</i>	Lauraceae	0.5207	genus	28.5	Try	zoo	Try	0.15	Goldenberg & Moraes, 2009
135	<i>Neoraputia alba</i>	Rutaceae	0.5604	family	16	Safar et al., 2022	nzoo	Safar et al., 2022	0.71	Safar et al., 2022
136	<i>Ocotea</i>	Lauraceae	0.5195	genus	16.09	Safar et al., 2022	zoo	Safar et al., 2022	1.12	Safar et al., 2022
137	<i>Ocotea aciphylla</i>	Lauraceae	0.5195	genus	19.95	Try	zoo	Safar et al., 2022	2	Brotto et al., 2013
138	<i>Ocotea argentea</i>	Lauraceae	0.5195	genus	18.18	Safar et al., 2022	zoo	Safar et al., 2022	0.9	Safar et al., 2022
139	<i>Ocotea confertiflora</i>	Lauraceae	0.5195	genus	14.79	Safar et al., 2022	zoo	Safar et al., 2022	0.91	Safar et al., 2022
140	<i>Ocotea indecora</i>	Lauraceae	0.5195	genus	19.06	Try	zoo	Safar et al., 2022	0.98	Safar et al., 2022

141	<i>Ocotea longifolia</i>	Lauraceae	0.5195	genus	20	Try	zoo	Safar et al., 2022	1.5	Safar et al., 2022
142	<i>Ocotea notata</i>	Lauraceae	0.5195	genus	5	REFLORA	zoo	Safar et al., 2022	0.6	Brotto et al., 2013
143	<i>Parapiptadenia pterosperma</i>	Fabaceae	0.7400	genus	20	Try	nzoo	Safar et al., 2022	1.4	Safar et al., 2022
144	<i>Pera glabrata</i>	Peraceae	0.6655	genus	24.7	Try	zoo	Safar et al., 2022	0.2	Safar et al., 2022
145	<i>Pera parvifolia</i>	Peraceae	0.6655	genus	10	IPÊ	zoo	Safar et al., 2022	0.2	Safar et al., 2022
146	<i>Persea</i>	Lauraceae	0.4508	genus	20	Try	zoo	Barbosa et al., 2016	5	Lorenzi, 2021
147	<i>Persea americana</i>	Lauraceae	0.4508	genus	20	Try	zoo	Barbosa et al., 2016	5	Lorenzi, 2021
148	<i>Piper cernuum</i>	PIPÉraceae	0.3300	genus	6	BDC	zoo	Try	0.08	Lobato et al., 2005
149	<i>Piptadenia gonoacantha</i>	Fabaceae	0.7467	genus	20	Lorenzi, 2021	nzoo	Try	1	REFLORA
150	<i>Piptadenia paniculata</i>	Fabaceae	0.7467	genus	34.2	Try	nzoo	Try	1.15	Safar et al., 2022
151	<i>Plathymenia reticulata</i>	Fabaceae	0.4850	genus	40	REFLORA	nzoo	Try	1.2	REFLORA
152	<i>Platypodium elegans</i>	Fabaceae	0.7500	genus	41.25	Try	nzoo	Try	1.5	REFLORA
153	<i>Pleroma estrellense</i>	Melastomataceae	0.5604	family	10	REFLORA	nzoo	REFLORA	1	REFLORA
154	<i>Pleroma fissinervia</i>	Melastomataceae	0.5604	family	8	REFLORA	nzoo	REFLORA	1	REFLORA
155	<i>Pourouma guianensis</i>	Urticaceae	0.3905	genus	28.75	Try	zoo	Barbosa et al., 2016	0.89	Safar et al., 2022
156	<i>Pouteria gardneri</i>	Sapotaceae	0.7583	genus	12	REFLORA	zoo	Try	1.2	REFLORA
157	<i>Protium heptaphyllum</i>	Burseraceae	0.5543	genus	20	Try	zoo	Try	1.5	REFLORA
158	<i>Pseudobombax grandiflorum</i>	Malvaceae	0.2925	genus	25	Lorenzi, 2021	nzoo	Try	3	REFLORA
159	<i>Pseudopiptadenia contorta</i>	Fabaceae	0.6449	genus	19.87	Safar et al., 2022	nzoo	Try	0.79	Safar et al., 2022
160	<i>Psidium guajava</i>	Myrtaceae	0.6845	genus	25	Try	zoo	Try	0.5	REFLORA
161	<i>Psidium myrtilloides</i>	Myrtaceae	0.6845	genus	20	REFLORA	zoo	Try	0.5	REFLORA
162	<i>Psychotria carthagenensis</i>	Rubiaceae	0.5200	genus	13.26	Safar et al., 2022	zoo	Try	0.43	Safar et al., 2022
163	<i>Psychotria pedunculosa</i>	Rubiaceae	0.5200	genus	5	REFLORA	zoo	Barbosa et al., 2016	0.4	FFESP
164	<i>Psychotria vellosiana</i>	Rubiaceae	0.5500	genus	6	IPÊ	zoo	Barbosa et al., 2016	0.3	FFESP
165	<i>Pterocarpus rohrii</i>	Fabaceae	0.4997	genus	50	Try	nzoo	Try	0.45	Safar et al., 2022
166	<i>Pterygota brasiliensis</i>	Malvaceae	0.5900	genus	30	Safar et al., 2022	nzoo	Safar et al., 2022	1.3	Safar et al., 2022
167	<i>Rauvolfia grandiflora</i>	Apocynaceae	0.5048	genus	25	Lorenzi, 2021	zoo	Lorenzi, 2021	3	Lorenzi, 2021
168	<i>Roupala montana</i>	Proteaceae	0.7967	genus	35	Try	nzoo	Try	1.1	Safar et al., 2022
169	Rubiaceae	Rubiaceae	0.5604	family	6	Safar et al., 2022	nzoo	Safar et al., 2022	0.1	Safar et al., 2022
170	<i>Sapium glandulosum</i>	Euphorbiaceae	0.4309	genus	33.81	Try	zoo	Try	0.6	Safar et al., 2022
171	<i>Schoepfia brasiliensis</i>	Schoepfiaceae	0.5604	SouthAmerica	19.73	Safar et al., 2022	zoo	Try	0.3	Safar et al., 2022
172	<i>Senefeldera verticillata</i>	Euphorbiaceae	0.7800	genus	16.5	Safar et al., 2022	nzoo	Safar et al., 2022	1	Safar et al., 2022
173	<i>Senegalia polyphylla</i>	Fabaceae	0.5604	family	17	Safar et al., 2022	nzoo	Try	0.6	Safar et al., 2022
174	<i>Senna macranthera</i>	Fabaceae	0.5555	genus	11.5	Try	nzoo	Barbosa et al., 2016	0.3	Pozitano & Rocha, 2011
175	<i>Senna multijuga</i>	Fabaceae	0.5555	genus	15	Try	nzoo	Barbosa et al., 2016	0.7	Dantas & Silva, 2013
176	<i>Serjania erecta</i>	Sapindaceae	0.5604	family	2	REFLORA	nzoo	Try	0.8	Silva et al., 2013
177	<i>Simarouba amara</i>	Simaroubaceae	0.3942	genus	32.5	Try	zoo	Try	0.8	Safar et al., 2022
178	<i>Siparuna guianensis</i>	Siparunaceae	0.6618	genus	10	REFLORA	zoo	Safar et al., 2022	0.4	Valentini et al., 2008
179	<i>Sloanea sinemariensis</i>	Elaeocarpaceae	0.7860	genus	30	Try	zoo	Safar et al., 2022	0.48	Safar et al., 2022
180	<i>Solanum</i>	Solanaceae	0.2800	genus	9	Safar et al., 2022	zoo	Try	0.5	REFLORA
181	<i>Solanum castaneum</i>	Solanaceae	0.2800	genus	3	Miranda, 2015	zoo	Barbosa et al., 2016	0.5	REFLORA
182	<i>Solanum leucodendron</i>	Solanaceae	0.2800	genus	20	Lafetá, 2002	zoo	Barbosa et al., 2016	0.5	REFLORA
183	<i>Solanum melissarum</i>	Solanaceae	0.2800	genus	4	REFLORA	zoo	Barbosa et al., 2016	0.5	REFLORA
184	<i>Solanum pseudoquina</i>	Solanaceae	0.2800	genus	13.6	Try	zoo	Try	0.33	Safar et al., 2022
185	<i>Sorocea guilleminiana</i>	Moraceae	0.5775	genus	20	Try	zoo	Try	0.88	Safar et al., 2022
186	<i>Sparattosperma leucanthum</i>	Bignoniaceae	0.5604	family	21.32	Safar et al., 2022	nzoo	Try	0.13	Safar et al., 2022
187	<i>Spondias macrocarpa</i>	Anacardiaceae	0.3945	genus	17.86	Safar et al., 2022	zoo	Barbosa et al., 2016	1.77	Safar et al., 2022
188	<i>Swartzia apetala</i>	Fabaceae	0.8407	family	13.06	Safar et al., 2022	zoo	Try	2	REFLORA

189	Swartzia linharensis	Fabaceae	0.8407	family	25	Safar et al., 2022	zoo	Safar et al., 2022	1.7	Safar et al., 2022
190	Swartzia myrtifolia	Fabaceae	0.8407	family	15.66	Safar et al., 2022	zoo	Safar et al., 2022	0.86	Safar et al., 2022
191	Swartzia simplex	Fabaceae	0.8407	family	35	Try	zoo	Safar et al., 2022	0.92	Safar et al., 2022
192	Syagrus pseudococos	Arecaceae	0.5604	family	6.75	Try	zoo	Try	1.91	Safar et al., 2022
193	Syzygium cumini	Myrtaceae	0.7000	genus	15	Try	zoo	Try	1.5	REFLORA
194	Syzygium jambos	Myrtaceae	0.7000	genus	15	Try	zoo	Try	2.5	REFLORA
195	Tachigali pilgeriana	Fabaceae	0.5834	genus	17.17	Safar et al., 2022	nzoo	Safar et al., 2022	1.55	Safar et al., 2022
196	Tachigali vulgaris	Fabaceae	0.5834	genus	13	Safar et al., 2022	nzoo	Try	1.5	Abreu et al, 2017
197	Tapirira guianensis	Anacardiaceae	0.3750	genus	35	Try	zoo	Try	0.7	Safar et al., 2022
198	Thyrsodium spruceanum	Anacardiaceae	0.5950	genus	14	Safar et al., 2022	zoo	Safar et al., 2022	1.26	Safar et al., 2022
199	Trema micrantha	Cannabaceae	0.2750	genus	39	Try	zoo	Try	0.3	Carvalho, 2003
200	Trembleya parviflora	Melastomataceae	0.5604	family	1.7	Try	nzoo	Try	0.5	Renato et al., 2015
201	Trichilia hirta	Meliaceae	0.6512	genus	38	Try	zoo	Try	0.6	REFLORA
202	Trichilia pallens	Meliaceae	0.6512	genus	20	Safar et al., 2022	zoo	Safar et al., 2022	0.61	Safar et al., 2022
203	Vernonanthura discolor	Asteraceae	0.5400	SouthAmerica	22.8	Try	nzoo	Try	0.3	Grzybowski et al., 2016
204	Vernonanthura divaricata	Asteraceae	0.5400	SouthAmerica	18	IPÊ	nzoo	Try	0.3	Grzybowski et al., 2016
205	Virola bicuhyba	Myristicaceae	0.4784	genus	28.5	Try	zoo	Try	2.7	Carvalho, 2003
206	Vismia brasiliensis	Hypericaceae	0.4637	genus	10	REFLORA	zoo	Try	0.2	Mourão & Beltrati, 2001
207	Vismia guianensis	Hypericaceae	0.4637	genus	13.1	Safar et al., 2022	zoo	Safar et al., 2022	0.11	Safar et al., 2022
208	Vismia martiana	Hypericaceae	0.4637	genus	13.33	Safar et al., 2022	zoo	Try	0.2	Mourão & Beltrati, 2001
209	Vitex polygama	Lamiaceae	0.5558	genus	12	Lorenzi, 2021	zoo	Try	1	Embrapa
210	Vochysia	Vochysiaceae	0.4441	genus	24	Safar et al., 2022	nzoo	Safar et al., 2022	0.57	Safar et al., 2022
211	Xylopia frutescens	Annonaceae	0.5790	genus	19	Try	zoo	Try	0.5	Safar et al., 2022
212	Xylopia sericea	Annonaceae	0.5790	genus	27.5	Try	zoo	Try	0.5	REFLORA
213	Zanthoxylum acuminatum	Rutaceae	0.5860	genus	20	REFLORA	zoo	Try	0.3	FFESP
214	Zanthoxylum rhoifolium	Rutaceae	0.5860	genus	17.54	Try	zoo	Safar et al., 2022	0.3	Safar et al., 2022
215	Zeyheria tuberculosa	Bignoniaceae	0.7700	genus	23	Lorenzi, 2021	nzoo	Try	6.2	Embrapa

Legend: BDC: Biblioteca Digital de Ciências, University of Campinas; Embrapa: Empresa Brasileira de Pesquisa Agropecuária; FFESP: Flora Fanerogâmica do Estado de São Paulo; IPÊ: Instituto de Pesquisas Ecológicas; REFLORA: REFLORA Programme; Try: TRY Plant Trait Database.

SUPPLEMENTARY MATERIAL

Abandoned Eucalyptus plantations can be used as catalysts for natural regeneration

Matheus C. Santos¹, Jeanpierre R. Mirano², Nathália V. H. Safar³, Luiz Fernando S. Magnago^{2*}

¹ Programa de Pós-Graduação em Ecologia e Conservação da Biodiversidade, Universidade Estadual de Santa Cruz, Ilhéus, Bahia, Brazil; ² Centro de Formação em Ciências e Tecnologias Agroflorestais, Universidade Federal do Sul da Bahia, Ilhéus, Bahia, Brazil; ³ Departamento de Botânica, Universidade Federal de Viçosa, Viçosa, Minas Gerais, Brazil.

* Corresponding author. E-mail: luiz_fsm@hotmail.com.

This Supplementary Material includes:

Figure S1. Phylogenetic tree of tree species sampled in mixed and native treatments, in the Atlantic Forest. The scale of this phylogenetic tree is in millions of years.

Figure S2. Linear regression between phylogenetic diversity and structure indices and species richness.

Figure S3. Plots of models of basal area of native species (BA) in relation to Mean Nearest Taxon Distance (MNTD) with outliers, in mixed and native treatments of natural regeneration in the Atlantic Forest. The red line represents the fitted curve and the gray areas are the 95% confidence intervals.

Table S1. Mean and standard deviation of phylogenetic metrics for mixed and native treatments. PD, MPD, and MNTD are expressed in millions of years. sesPD, NRI, and NTI are expressed in units of standard deviation.

Table S2. Values of diversity indices and phylogenetic structure in mixed and native treatments.

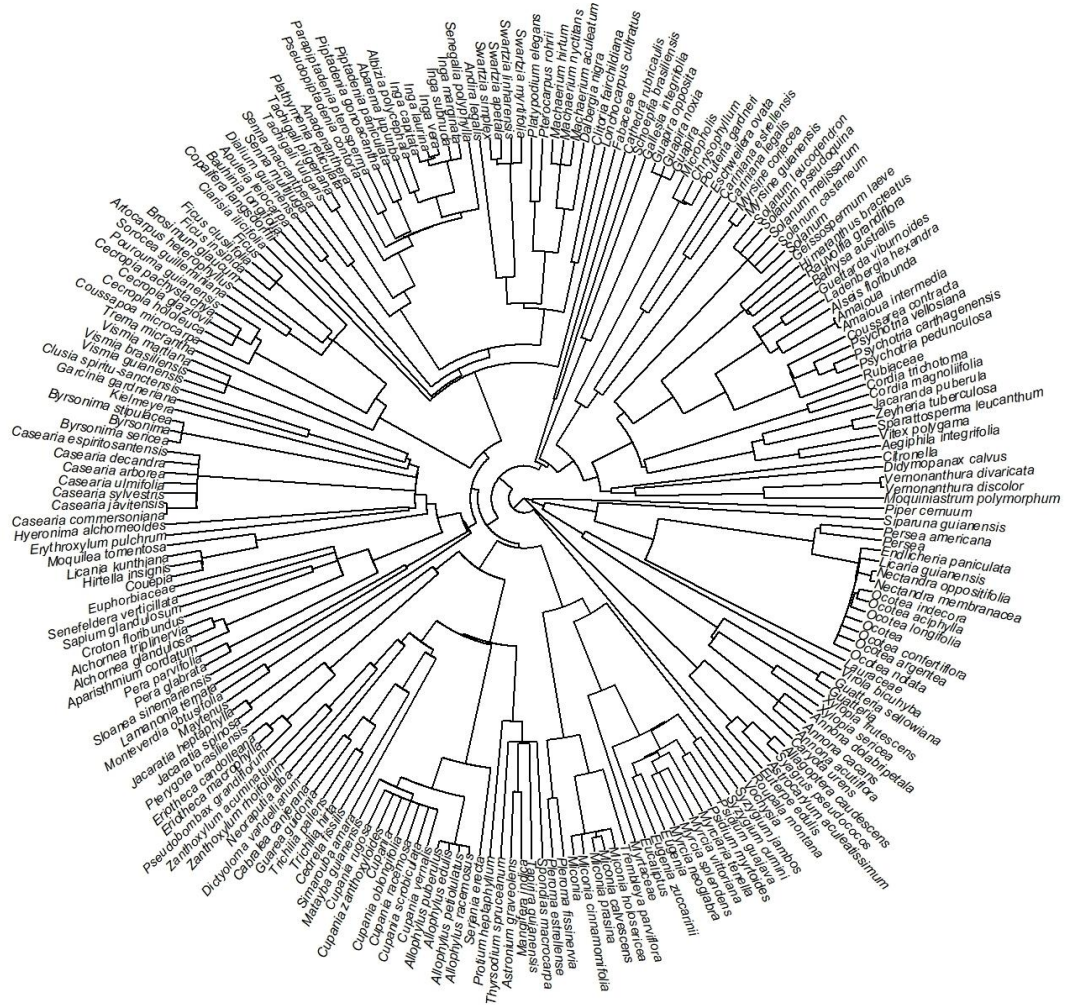


Figure S1. Phylogenetic tree of tree species sampled in mixed and native treatments, in the Atlantic Forest. The scale of this phylogenetic tree is in millions of years.

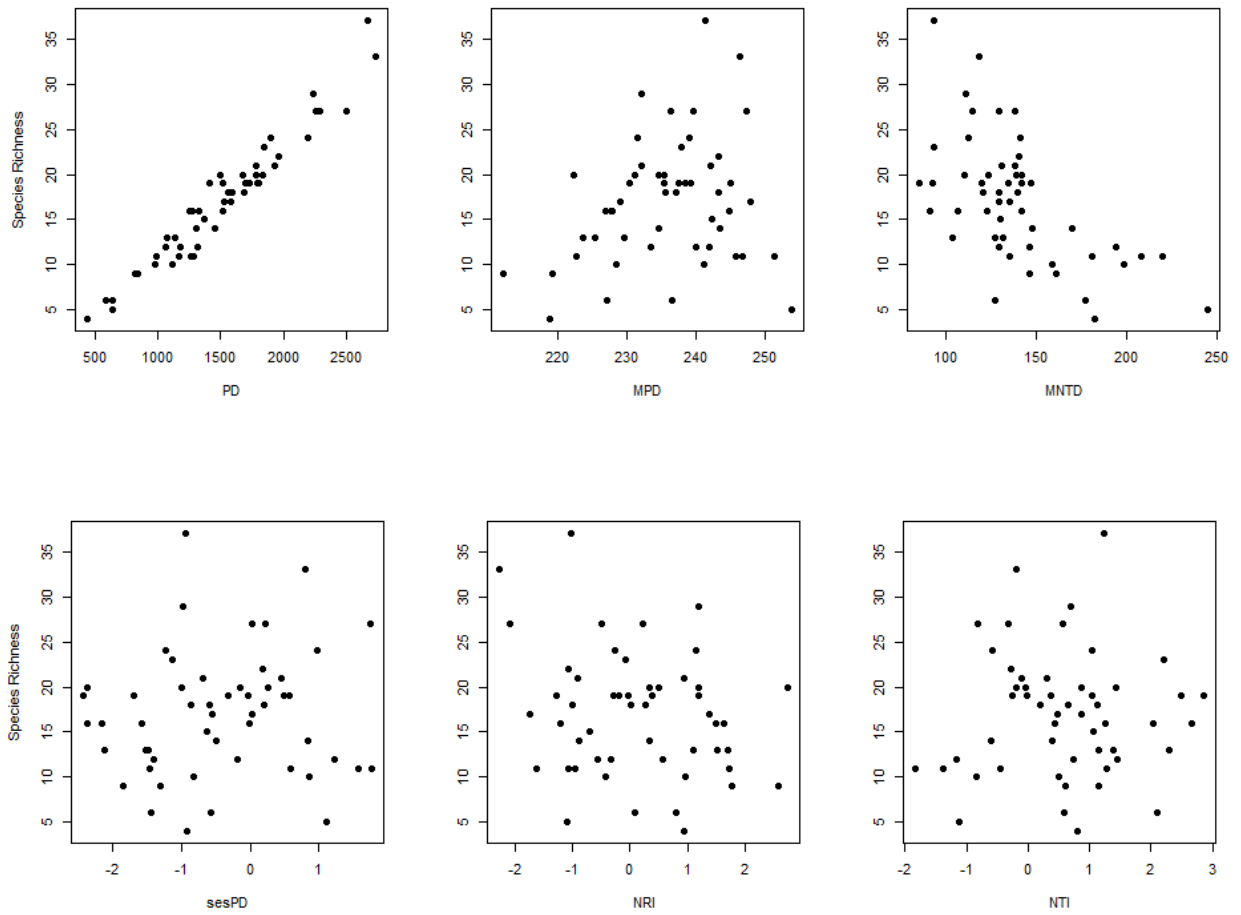


Figure S2. Linear regression between phylogenetic diversity and structure indices and species richness.

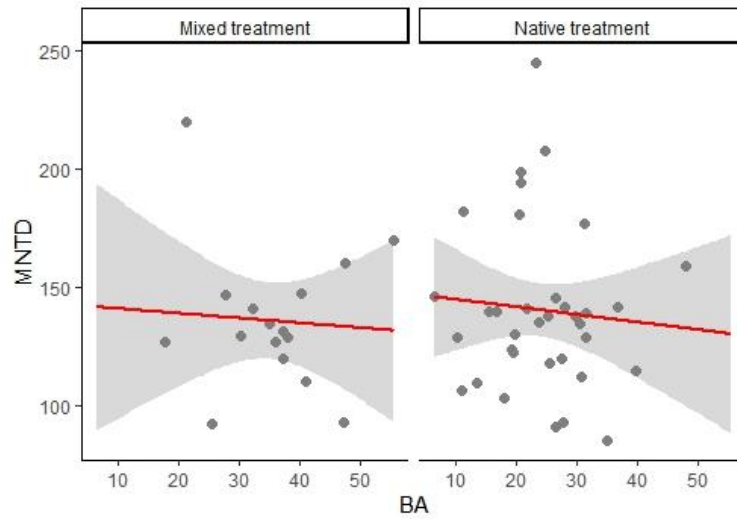


Figure S3. Plots of models of basal area of native species (BA) in relation to Mean Nearest Taxon Distance (MNTD) with outliers, in mixed and native treatments of natural regeneration in the Atlantic Forest. The red line represents the fitted curve and the gray areas are the 95% confidence intervals.

Table S1. Mean and standard deviation of phylogenetic metrics for mixed and native treatments. PD, MPD, and MNTD are expressed in millions of years. sesPD, NRI, and NTI are expressed in units of standard deviation.

Phylogenetic metric indices	Treatment		Student's t-test	
	Mixed	Native	t	P-value
PD	1436.02 ± 438.93	1534.29 ± 555.60	-0.692	0.49
MPD	235.10 ± 8.27	235.89 ± 9.25	-0.313	0.75
MNTD	135.98 ± 29.51	140.44 ± 34.88	^a	0.78
sesPD	-0.55 ± 1.02	-0.39 ± 1.15	-0.503	0.61
NRI	0.23 ± 1.04	0.12 ± 1.26	0.334	0.74
NTI	0.75 ± 1.10	0.49 ± 1.03	0.800	0.42

^a The data were not normal (Mann-Whitney U test: $W = 312$). **Legend:** PD: Phylogenetic Diversity; MPD: Mean Pairwise Distance; MNTD: Mean Nearest taxon distance; sesPD: standardized effect size of PD; NRI: Net Relatedness Index; NTI: Nearest Taxon Index.

Table S2. Values of diversity indices and phylogenetic structure in mixed and native treatments.

Transect	Treatment	SR	PD	MPD	MNTD	sesPD	pd.obs.p	NRI	NTI
1	Mixed	6	586.662	236.445	127.402	-1.438	0.082	0.092	2.103
2	Mixed	9	842.729	219.120	160.614	-1.300	0.115	1.758	0.618
3	Mixed	13	1134.418	225.303	131.564	-1.509	0.068	1.520	1.145
4	Mixed	18	1558.575	237.083	120.202	-0.865	0.193	0.023	1.134
5	Mixed	11	986.326	222.715	134.862	-1.444	0.079	1.714	1.272
6	Mixed	11	1281.748	246.586	219.597	1.764	0.978	-1.071	-1.830
7	Native	10	1110.426	241.151	198.621	0.859	0.799	-0.434	-0.843
8	Mixed	15	1372.634	242.185	129.944	-0.618	0.262	-0.691	1.061
9	Native	4	436.472	218.765	181.981	-0.906	0.172	0.930	0.791
10	Native	6	638.169	227.038	176.974	-0.576	0.260	0.791	0.586
11	Native	16	1327.441	226.808	122.449	-1.572	0.075	1.621	1.262
12	Native	13	1069.893	229.486	103.374	-2.110	0.023	1.106	2.293
13	Native	18	1687.563	243.232	139.786	0.202	0.562	-0.992	0.205
14	Native	12	1314.705	239.970	194.272	1.214	0.901	-0.323	-1.173
15	Native	20	1784.872	234.636	139.162	-0.137	0.438	0.488	-0.031
16	Native	12	1180.059	241.789	145.888	-0.176	0.420	-0.563	0.728
17	Native	20	1675.553	231.005	123.721	-0.989	0.159	1.195	0.876
18	Mixed	14	1449.002	243.369	169.934	0.838	0.791	-0.883	-0.594
19	Mixed	19	1793.399	238.373	146.778	0.486	0.664	-0.184	-0.257
20	Mixed	23	1845.954	237.755	93.511	-1.126	0.148	-0.081	2.195
21	Mixed	19	1517.623	235.237	92.320	-1.682	0.043	0.393	2.488
22	Mixed	13	1132.563	223.669	127.205	-1.463	0.072	1.704	1.379
23	Native	16	1271.343	227.710	91.145	-2.147	0.025	1.498	2.655
24	Native	19	1413.743	230.333	85.215	-2.412	0.016	1.182	2.858
25	Native	20	1833.220	235.326	141.729	0.254	0.598	0.334	-0.192
26	Mixed	18	1586.952	235.456	129.465	-0.578	0.282	0.260	0.656
27	Mixed	14	1308.406	234.494	147.674	-0.493	0.320	0.330	0.395
28	Native	27	2294.825	236.329	128.907	0.223	0.599	0.220	-0.332
29	Native	37	2668.021	241.247	93.084	-0.924	0.182	-1.021	1.240
30	Mixed	17	1583.126	247.869	129.035	0.038	0.482	-1.739	0.864
31	Mixed	24	2197.984	238.968	141.151	0.978	0.829	-0.272	-0.573
32	Native	19	1696.460	239.248	119.833	-0.323	0.359	-0.296	1.032
33	Native	19	1726.471	237.513	134.578	-0.029	0.479	-0.031	0.360
34	Native	17	1523.515	228.977	135.294	-0.555	0.276	1.366	0.487
35	Native	27	2254.402	239.498	114.820	0.030	0.505	-0.489	0.557
36	Native	19	1800.001	244.879	141.612	0.576	0.718	-1.288	-0.010
37	Mixed	29	2234.281	232.115	110.552	-0.975	0.161	1.178	0.694
38	Native	21	1926.568	242.102	138.169	0.452	0.663	-0.917	-0.115
39	Native	11	1264.257	251.273	207.613	1.568	0.964	-1.629	-1.384
40	Native	9	812.372	212.086	146.179	-1.832	0.049	2.570	1.145
41	Native	10	971.997	228.434	158.905	-0.814	0.211	0.961	0.500
42	Native	16	1514.645	244.806	141.438	-0.016	0.479	-1.213	0.444
43	Native	11	1170.749	245.660	181.092	0.592	0.711	-0.958	-0.447
44	Native	24	1896.924	231.373	112.170	-1.224	0.115	1.140	1.040
45	Native	16	1254.341	227.828	106.750	-2.361	0.020	1.488	2.023
46	Native	22	1966.193	243.162	139.991	0.185	0.562	-1.083	-0.283
47	Native	12	1064.522	233.322	129.172	-1.397	0.090	0.566	1.439
48	Native	33	2736.877	246.217	118.455	0.793	0.788	-2.268	-0.200
49	Native	21	1785.579	232.076	130.477	-0.685	0.248	0.931	0.296
50	Native	20	1493.563	222.194	109.927	-2.359	0.006	2.740	1.437
51	Native	5	631.130	253.800	244.885	1.099	0.925	-1.084	-1.130
52	Native	27	2503.590	247.209	137.816	1.735	0.968	-2.077	-0.828

Legend: SR: Species richness; PD: Phylogenetic Diversity; MPD: Mean Pairwise Distance; MNTD: Mean Nearest taxon distance; sesPD: standardized effect size of PD; NRI: Net Relatedness Index; NTI: Nearest Taxon Index.